



kinetics is well described by a pseudo-second order model. The aerogels with intermediate and high contact angles show higher adsorption capacities, with favourable isotherms, for the phenolic compounds with higher hydrophobicity, which shows the important role of hydrophobic interactions in this process. The best results were obtained for the aerogel functionalized with  $\beta$ -cyclodextrin when adsorbing the 4-chlorophenol, proving the positive effect of the presence of the  $\beta$ -cyclodextrin hydrophobic cavity in the adsorption of this highly hydrophobic pollutant.

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## 1. Introduction

The presence of phenolic compounds in many industrial wastewaters, such as from petrochemical, pharmaceutical/cosmetics, polymers, paints/dyes, cork and textiles industries, among others, is of concern in terms of environmental/human health protection. Even in low concentrations, these compounds present high toxicity and low biodegradability and, as they also exhibit high volatility, they easily spread and migrate. More complex structures (higher degree of substitution in the aromatic ring) usually give rise to increased toxicity (e.g., alkylphenols, chlorophenols, bisphenol-A). The removal of phenolic compounds from wastewaters is also crucial to comply with the International Legislation, being their adsorption a conventional and effective route for industrial effluents, usually using activated carbon [1–3]. Adsorbents with high selectivity and adsorption capacity are needed, and the silica-based aerogels are promising materials and have attracted great interest for this purpose due to their extremely high porosity/surface area, chemical versatility and possibility of reutilization [4–7].

As the phenolic compounds have functional groups of hydrophobic and hydrophilic character, several types of interactions with the adsorbents are expected, namely hydrogen bonding, hydrophobic and electrostatic interactions, being these interactions also dependent on the pH of the solution, in particular the latter one [8–11]. Thus, an improved adsorption may be achieved by tailoring the aerogels surface chemistry. In addition, the aerogels characteristic extensive network of micro and mesopores [6] greatly favours the adsorption process.

The tailoring of the chemistry of silica-based aerogels may be performed by the co-precursor method [6,12], by co-condensation of silica precursors that give rise to inherently hydrophilic silicas, such as tetramethylorthosilicate (TMOS) and tetraethylorthosilicate (TEOS), with organosilanes that offer the possibility of compounding the silica network with hydrophobic groups. Examples of the later are alkyltrialkoxysilanes, such as methyltrimethoxysilane (MTMS), methyltriethoxysilane (MTES), vinyltriethoxysilane (VTES), and many others. Another possible method is the surface modification/derivatization of the gel [6,12], in which the silica native gel (hydrophilic) is obtained first and then kept in a bath containing a mixture of solvent and surface modifying agent/silylating agent (usually organosilanes) that will condensate in the gel surface. In this method, the mass transfer in the gel takes place by diffusion only. Therefore, large amounts of solvent and significant time are required to achieve solvent exchange and subsequent surface modification, respectively, being thus very costly.

Regarding to the adsorption of phenolic compounds on silica-based materials, including aerogels/xerogels, only a few studies were found in the literature. Wu et al. [9–11] compared the adsorption of organic dyes (alizarin and phenol red) on several xerogels, synthesized by the co-precursor approach combining TEOS with several triethoxysilanes and, also, with bis(trimethoxysilyl)hexane (BTMSH) and bis(trimethoxysilylpropyl)amine (BTMSPA). They concluded that the organic dyes adsorption in the xerogels is influenced by all types of interactions referred above, but the hydrophobic interactions are of major relevance. The adsorption

capacity of phenol red was the lowest among all the tested adsorbents (always below  $45 \text{ mg g}^{-1}$ ). You et al. [13] also tested a xerogel made from TEOS/BTMSH system, but for adsorption of bromophenol blue. The obtained adsorption capacity did not reach values above  $20 \text{ mg g}^{-1}$ . Haghbeen and Legge [14] studied the combination of TMOS and TEOS with propyltrimethoxysilane (PTMS) and methyltriethoxysilane (MTPS) to produce xerogel adsorbents for many types of phenolic compounds, but the adsorption capacities were not provided.

Concerning the surface derivatization method, An et al. have tested silica particles as adsorbents of phenol after performing the grafting of polyethyleneimine (PEI) [15,16] and polymethacrylic acid (PMAA) [8] onto the particles surface. They obtained a high adsorption capacity for phenol ( $160 \text{ mg g}^{-1}$  and  $163 \text{ mg g}^{-1}$  for PEI and PMAA, respectively) justified by the hydrogen bonding and electrostatic interactions. Faraji et al. [17,18] performed the modification of the surface of silica gel particles by compounding it with  $\beta$ -cyclodextrin, using the linking agent 3-glycidoxypropyltrimethoxysilane, and tested this adsorbent for phenol and many phenolic compounds with different types of functional substituent groups in the aromatic ring. Although it was referred that good adsorption performance was achieved, no numeric data was provided. Qin et al. [7] have modified the surface of a TEOS-derived xerogel with TMCS and got an adsorption capacity of  $142 \text{ mg g}^{-1}$  for phenol. Finally, Anbia and co-workers [19,20] had used mesoporous silica (MCM-41 and SBA-15) and made its surface derivatization with aminopropyltrimethoxysilane, in order to obtain better adsorption capacity for chlorophenols, bromophenols and iodophenols. They achieved very good adsorption performances and the best result was obtained for modified MCM-41, which showed an adsorption capacity of  $339 \text{ mg g}^{-1}$  for 2,4,6-trichlorophenol.

The literature results described above allow concluding that the hydrophobic/hydrophilic interactions between the silica-based adsorbent and the phenolic compounds are complex and very important to the adsorption capacity of the adsorbents. In an earlier work [4], we had tested MTMS-derived aerogels and xerogels, which were highly porous, flexible and hydrophobic [21–23], to the adsorption of the aromatic solvents benzene, toluene and phenol from water. The higher adsorption capacities were obtained for the aerogel in the cases of toluene ( $173 \text{ mg g}^{-1}$ ) and benzene ( $192 \text{ mg g}^{-1}$ ), which were remarkably high when compared to the available literature data. However, for phenol the adsorption performance was not the best. Thus, we have now focused on the chemical modification of the aerogels/xerogels obtained from MTMS, in order to improve their adsorption capacity towards phenolic compounds.

In this work, the co-precursor method is applied to obtain aerogels and xerogels of tailored hydrophobicity/hydrophilicity, by combining different proportions of MTMS and TMOS which contribute to have a balance of  $-\text{CH}_3$  and  $-\text{OH}$  in the final structure of the gel. In addition,  $\beta$ -cyclodextrin ( $\beta$ -CD) was added along with the precursors to one of the formulations in order to create a dispersion of this macromolecule in the gel matrix. It is recognized that the hydrophobic hole of  $\beta$ -CD can act as a cage for the aromatic ring of phenol. With these chemically tailored materials, the aim is to

investigate their adsorption interactions with three phenolic compounds, phenol, cresol and 4-chlorophenol, which show increasing hydrophobic character, and with one surfactant, SDBS, with a structure somewhat similar to phenolic compounds. Equilibrium and kinetic studies were carried out for this purpose. The understanding of the nature of dominant adsorption interactions of phenolic compounds with the adsorbents is a crucial step to design more selective materials for the specific moieties that these compounds can present.

## 2. Materials and methods

### 2.1. Materials

For the synthesis of aerogels/xerogels, TMOS ( $\geq 98$ ), MTMS ( $\geq 98\%$ ), ammonium hydroxide (25%  $\text{NH}_3$  in  $\text{H}_2\text{O}$ ), methanol ( $\geq 99.8\%$ , MeOH) and  $\beta$ -cyclodextrin ( $\geq 97\%$ ,  $\beta$ -CD) were purchased from Sigma–Aldrich. Ethanol ( $\geq 99.8\%$ , EtOH) was purchased from Fisher. All reagents were used without further purification.

To prepare the synthetic effluents of this work, analytical grade phenol (99%), *p*-cresol (99%), 4-chlorophenol (99%) and sodium dodecylbenzenesulfonate (80%, SDBS) were obtained from Aldrich. Deionised water of high purity (from a Milipore ultra-pore water system) was used.

The structures and some chemical properties of the used precursors/additives, selected pollutants and SDBS are present in Table 1.

Before the adsorption tests, both aerogels and xerogels adsorbents were ground and passed through two sieves to retain the particle sizes in the interval 74–250  $\mu\text{m}$ .

### 2.2. Synthesis of adapted silica-based aerogels and xerogels

#### 2.2.1. Aerogels/xerogels synthesized from TMOS–MTMS chemical system

MTMS and TMOS precursors were diluted in MeOH and a basic solution of  $\text{NH}_4\text{OH}$  (1M) was added. The mixture was stirred during 10 min and the resultant sol was transferred to plastic syringes and aged during five days at 27 °C. The molar ratios of the system components were: MTMS:TMOS – (0.2–0.8) to (0.8–0.2); Si:MeOH – 12; Si: $\text{H}_2\text{O}$  – 4. The obtained gels were dried by two drying methods: ambient pressure drying (APD) and supercritical fluids extraction (SFE). The APD method consists in subjecting the samples to several temperature cycles in an oven. The used temperatures were 60 °C (24 h), 100 °C (1 h), 150 °C (1 h) and 200 °C (1 h). In the SFE method, the gel was firstly washed with the solvent and, then, a continuous flow of supercritical  $\text{CO}_2$  was passed in the sample to promote its drying.

#### 2.2.2. Aerogels synthesized from TMOS–MTMS– $\beta$ -CD chemical system

The synthesis of the aerogels with  $\beta$ -CD was slightly different than described in Section 2.2.1, in order to overcome the  $\beta$ -CD solubility limitation. Initially 0.18 g of  $\beta$ -CD was dissolved in a solution containing a solvent mixture of  $\text{H}_2\text{O}$ , EtOH (1:2.3 v/v) and  $\text{NH}_4\text{OH}$ . Then the precursors (TMOS and MTMS) were added and the resultant solution was stirred during 10 min and kept in an oven at 27 °C, during 5 days, for gelation. The used drying method was SFE, already described in Section 2.2.1. The molar ratios of MTMS:TMOS:MeOH: $\text{H}_2\text{O}$ : $\beta$ -CD were 0.5:0.5:12:4:2.64  $\times 10^{-3}$ .

For simplicity, the aerogels and xerogels of this work were identified by a short designation, as described in the following two examples. Sample A-50M50T-CD corresponds to an Aerogel (A), with 50% of the Si from MTMS (50M) and 50% of the Si from TMOS (50T), and  $\beta$ -CD was added in the system (CD); sample X-50M50T means that it is a xerogel (X), with 50% of the Si from both MTMS (50M) and TMOS (50T).

### 2.3. Aerogels/xerogels characterization methods

#### 2.3.1. FTIR spectroscopy

FTIR spectroscopy was performed on a Jasco FT/IR 4200 spectrometer. The spectra were obtained in the KBr mode, using KBr pellets composed by 80 mg of KBr and 0.2–0.3 mg of aerogel/xerogel, in a wave number range of 400–4000  $\text{cm}^{-1}$ .

#### 2.3.2. Contact angle measurement

The contact angle of the materials was obtained by dropping water droplets on the surface of pre-compressed pellets of aerogel/xerogel, in order to obtain the hydrophobicity/hydrophilicity level of each sample. The used equipment and software for the contact angle evaluation was from Dataphysics, model OCA 20.

#### 2.3.3. Bulk and skeletal densities evaluation

By helium pycnometry (Micromeritics, Accupyc 1330), it was possible to measure the skeletal density of the aerogels/xerogels, which along with the bulk density allows obtaining the porosity of the materials. The bulk density was obtained by weighting the samples and measuring their volume by immersion in ethanol.

#### 2.3.4. Scanning electron microscopy

The microstructure of the aerogels/xerogels was observed by scanning electron microscopy (SEM), using a microscope from JOEL, model JMS-5310. The samples were previously coated with gold to overcome their low electric conductivity.

#### 2.3.5. Surface area evaluation and porosimetry

The specific surface area and pore size distribution/average pore size were obtained by nitrogen gas adsorption, with the BET adsorption theory and the BJH desorption method, respectively, and using ASAP 2000 equipment, from Micromeritics. Previous degasification of the samples was performed in a vacuum oven at 50 °C, during 3 days, and also in the equipment.

#### 2.3.6. Simultaneous differential thermal analysis

A Simultaneous Differential Thermal Analyzer (DSC/TGA), model SDT Q500, from TA Instruments, was used to observe the thermal behaviour of the aerogels and to obtain confirmation of the presence of  $\beta$ -CD in the samples. The samples were placed in an alumina pan and heated from room temperature to 1200 °C, at a constant heating rate of 10 °C/min, under a nitrogen flow.

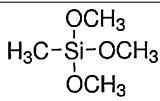
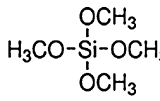
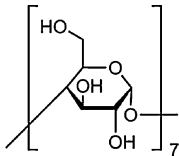
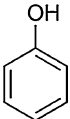
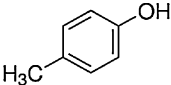
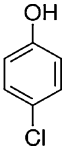
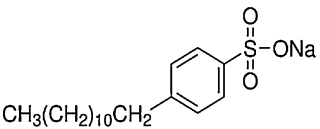
### 2.4. Adsorption tests with SDBS and phenolic compounds

Concentrations of SDBS and phenolic compounds were determined by using the UV–Vis spectrophotometer T70, from PG Instruments. The concentrations used in the calibration curves were in the range of 1–60  $\text{mg L}^{-1}$  for phenol and *p*-cresol, 1–20  $\text{mg L}^{-1}$  for 4-chlorophenol and 0.5–40  $\text{mg L}^{-1}$  for SDBS. The solutions were analyzed by measuring the absorbance at 270 nm (phenol), 276 nm (*p*-cresol), 280 nm (4-chlorophenol) and 222 nm (SDBS). Quartz glass cells (Hellma) of 10 mm path length were used.

#### 2.4.1. Adsorption equilibrium and kinetics

Equilibrium isotherms for the system X-50M50T/SDBS were determined through batch tests, where 45 mL of SDBS solutions with several concentrations (87, 174, 348, 1742, 3485 and 6970  $\text{mg L}^{-1}$ ) were added into glass bottles (50 mL) containing an appropriate amount of adsorbent (0.09 g). These flasks were agitated at 16 rpm in a shaker (Heidolph – REAX 20) at room temperature, and samples for analysis were withdrawn after 24 h of

**Table 1**  
Properties of selected precursors and phenol-derivative compounds.

Precursors/additives					
Compound	Chemical structure	MW (g mol <sup>-1</sup> )	S(H <sub>2</sub> O) <sup>a</sup> (g L <sup>-1</sup> ) [24]	Log P <sub>ow</sub> <sup>b</sup>	
Methyltrimethoxysilane (MTMS)		136.22	80; hydrolysis	0.7 [24]	
Tetramethyl orthosilicate (TMOS)		152.22	Hydrolysis; soluble in organic solvents	–	
$\beta$ -Cyclodextrin ( $\beta$ -CD)		1134.98	14.3	–	
Phenol-derivative compounds					
Compound	Chemical structure	MW (g mol <sup>-1</sup> )	S(H <sub>2</sub> O) <sup>c</sup> (g L <sup>-1</sup> ) [25]	Log P <sub>ow</sub> <sup>b</sup>	LD <sub>50</sub> (rats) (mg g <sup>-1</sup> ) [24]
Phenol		94.11	94.8	1.5 [25]	317
<i>p</i> -Cresol		104.14	21.52	1.9 [26]	207
4-Chlorophenol		128.56	28	2.4 [26]	670
Sodium dodecylbenzene sulfonate (SDBS) <sup>d</sup>		348.48	200	0.45 [27]	438

<sup>a</sup> Solubility in water at 20 °C.<sup>b</sup> Log<sub>10</sub> partition coefficient *n*-octanol/water at 25 °C.<sup>c</sup> Solubility in water at 25 °C.<sup>d</sup> Critical micelle concentration (CMC) is 1.2 × 10<sup>-3</sup> M (418.2 g L<sup>-1</sup>) [28].

contact time in order to determine the concentration of SDBS present in solution.

Equilibrium adsorption curves of phenol, *p*-cresol and 4-chlorophenol onto X-50M50T, A-20M80T, A-50M50T, A-80M20T and A-50M50T-CD aerogels/xerogels were obtained by following the same procedure, but with different initial concentrations of adsorbate, varying between 10 and 1000 mg L<sup>-1</sup>. For each concentration of adsorbate (SDBS or phenolic compounds), two replicas were prepared.

The adsorbate uptake ( $q_e$ ), expressed as adsorbate removal per unit mass of adsorbent (mg g<sup>-1</sup>), was calculated according to Eq. (1), where  $C_0$  is the initial adsorbate concentration (mg L<sup>-1</sup>),  $C_e$  the equilibrium concentration (mg L<sup>-1</sup>),  $V$  the batch volume (L) and  $m$  the adsorbent mass (g). The percentage of adsorbate removed from the aqueous solution was calculated using Eq. (2), where  $C_0$  (mg L<sup>-1</sup>) and  $C_e$  (mg L<sup>-1</sup>) are the initial and equilibrium adsorbate concentrations, respectively.

$$q_e = \frac{(C_0 - C_e)V}{m} \quad (1)$$

$$R (\%) = \frac{C_0 - C_e}{C_0} \times 100 \quad (2)$$

Preliminary kinetic tests were carried out in order to determine the minimum contact time to establish equilibrium conditions between X-50M50T and SDBS. In these tests, 0.09 g of adsorbent and 45 mL of adsorbate solution with initial concentration 200 mg L<sup>-1</sup> were poured into 50 mL flasks. These experiments were carried out under the same conditions as those used in the adsorption equilibrium experiments (temperature and agitation), and samples for analysis were collected after a certain contact time that varied between 5 and 1440 min. After establishing the contact time (360 min), experiments were performed in order to determine the SDBS uptake along time, varying the feed solution concentrations

(11, 22, 44 and 87 mg L<sup>-1</sup>) at some conditions of temperature, adsorbent amount and adsorbate volume described above.

#### 2.4.2. Kinetic models

The kinetics of SDBS adsorption was evaluated applying two different models: (1) the pseudo-first-order kinetic model [29], and (2) pseudo-second-order kinetic model [30].

The linear form of the equation for the pseudo-first-order is expressed as:

$$\ln(q_e - q_t) = \ln(q_e) - k_1 t \quad (3)$$

where  $q_e$  and  $q_t$  are the amounts of SDBS adsorbed on the adsorbent (mg g<sup>-1</sup>) at equilibrium and at time  $t$ , respectively, and  $k_1$  is the Lagergren pseudo-first order adsorption rate constant (min<sup>-1</sup>).

The linear form of the pseudo-second-order model is written as:

$$\frac{t}{q_t} = \frac{1}{k_2 \cdot q_e^2} + \frac{1}{q_e} \cdot t \quad (4)$$

where  $k_2$  is the second-order adsorption rate constant (g mg<sup>-1</sup> min<sup>-1</sup>).

#### 2.4.3. Adsorption isotherm models

The data correlation either using theoretical or empirical models is essential for the interpretation of experimental adsorption isotherms. The determination of the adsorption capacity at various equilibrium concentrations ( $C_e$ ) was performed by obtaining the experimental adsorption isotherm. The commonly used Langmuir and/or Freundlich models were fitted to the experimental data. These models are given by Eqs. (5) and (6), respectively.

$$q_e = \frac{q_{\max} K C_e}{1 + K C_e} \quad (5)$$

$$q_e = K_F \cdot C_e^{1/n} \quad (6)$$

Langmuir parameters  $q_{\max}$  (mg L<sup>-1</sup>) and  $K$  (mg L<sup>-1</sup>) of Eq. (5) are the maximum capacity of adsorption at high equilibrium concentrations (monolayer of adsorbate) and the equilibrium constant, respectively. The parameters  $K_F$  (mg g<sup>-1</sup> (mg L<sup>-1</sup>)<sup>1/n</sup>) and  $1/n$  of Eq. (6) are the Freundlich capacity factor and the Freundlich intensity parameter, respectively [31]. The concentration of adsorbate retained in the adsorbent phase,  $q_e$  (mg g<sup>-1</sup>), and the percentage of adsorbate removed from aqueous solution were calculated by Eqs. (1) and (2) defined in Section 2.4.1.

### 3. Results and discussion

#### 3.1. Adsorbents characterization

##### 3.1.1. Chemical structure and hydrophilicity/hydrophobicity of the adsorbents

Fig. 1 presents the FTIR spectra of the xerogel and aerogels synthesized with the system MTMS-TMOS. The ascribing of the vibration bands to the chemical bonds, also presented in this figure, considered the characteristic vibration frequencies of organic and silica-based materials published in the literature [32,33].

The characteristic broad band/double band due to the asymmetric stretching vibrations of siloxane bonds (Si–O–Si) appears at ~1100 cm<sup>-1</sup> and ~1270 cm<sup>-1</sup>, and the double band seen for the sample A-80M20T transforms gradually in a larger band with a shoulder (less defined band at 1270 cm<sup>-1</sup>) with the increasing amount of TMOS in the precursor system. This was expected, as the pattern observed for the sample A-20M80T is similar to those obtained for native silica aerogels, which exhibit a higher degree of structural order when compared to MTMS-derived aerogels. The symmetric stretching vibrations of the same bonds are observed in the sharp peak near 780 cm<sup>-1</sup>, and this peak become broader, less

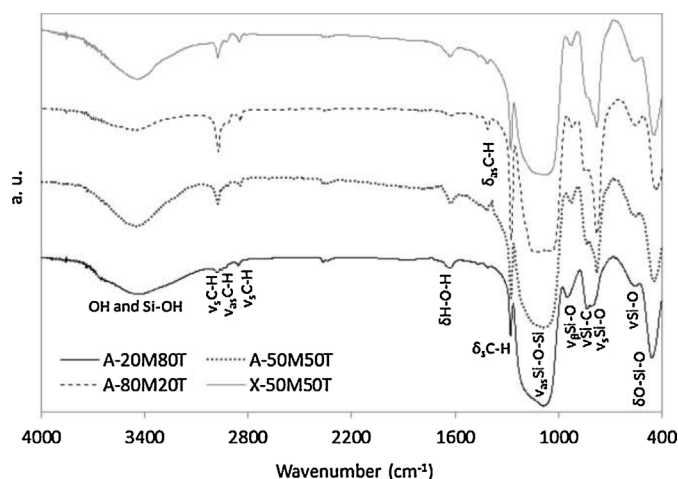


Fig. 1. FTIR spectra for MTMS-TMOS-derived xerogel and aerogels ( $\nu$ , stretching vibration;  $\nu_s$ , symmetric stretching vibration;  $\nu_{as}$ , asymmetric stretching vibration;  $\delta$ , deformation vibration;  $\delta_s$ , symmetric deformation vibration;  $\delta_{as}$ , asymmetric deformation vibration (bending);  $\nu_B$ , in plane-stretching vibration).

intense and shifts to higher wavenumbers with increasing amount of TMOS. Finally, the peak corresponding to the deformation vibration of O–Si–O bonds appears around 450 cm<sup>-1</sup>; the small broad bands seen at 920–950 cm<sup>-1</sup> and at 550 cm<sup>-1</sup> are related to the stretching vibrations in-plane of Si–O bonds and to the stretching vibrations of the same bonds in silica defects, respectively.

The stretching vibrations of Si–C bonds appear at ~840 cm<sup>-1</sup> and become less intense with a smaller amount of MTMS in the precursor system, as expected. The symmetric and asymmetric stretching vibrations of the C–H bonds of the –CH<sub>3</sub> groups linked to Si show between 2840 and 3000 cm<sup>-1</sup>, also vanishing with decreasing amount of MTMS. The symmetric deformation vibrations of the C–H bonds are visible in the sharp peak at 1410 cm<sup>-1</sup>, being the asymmetric mode also observed, at 1460 cm<sup>-1</sup>.

Hydroxyl groups are also present in the structure, noted by the broad bands near 1625 cm<sup>-1</sup> and in the range 3200–3600 cm<sup>-1</sup>, and these bands are more evident with high amounts of TMOS in the precursor system (samples A-50M50T and A-20M80T), which was also expected since TMOS gives rise to more OH terminal groups in the structure.

In summary, these spectra prove that obtained adsorbents are mainly composed by a silica backbone, with methyl groups linked to one part of the Si atoms and OH terminal groups. The chemical structure exhibited by the xerogel (X-50M50T) is similar to that of the aerogel counterpart. The structural order of the silica network increases with the relative amount of TMOS/MTMS, which is seen by the shifting and shape changing of some bands. The intensity of the Si–CH<sub>3</sub> and Si–OH groups also changes with the proportion of the precursors, as expected, proving that the gels surface is being effectively modified by this precursor combination.

For the aerogel synthesized with  $\beta$ -CD, the characteristic FTIR peaks of this macromolecule are masked by the larger bands of silica until 1400 cm<sup>-1</sup>. Thus, it was decided to perform TGA, in order to confirm the presence of  $\beta$ -CD in the structure. The obtained TGA curves are presented in Fig. 2. It is possible to verify that  $\beta$ -CD thermally decomposes between 314 and 332 °C. For the sample A-50M50T-CD a weight loss of 2.1% is observed in a similar interval (304–346 °C), and this is certainly due to the decomposition of  $\beta$ -CD since it does not occur for the sample A-50M50T. The weight losses that starts at 370 °C for samples A-50M50T and A-50M50T-CD are due to the decomposition of the methyl groups of the aerogels structure [23], a phenomenon that changes significantly the contact

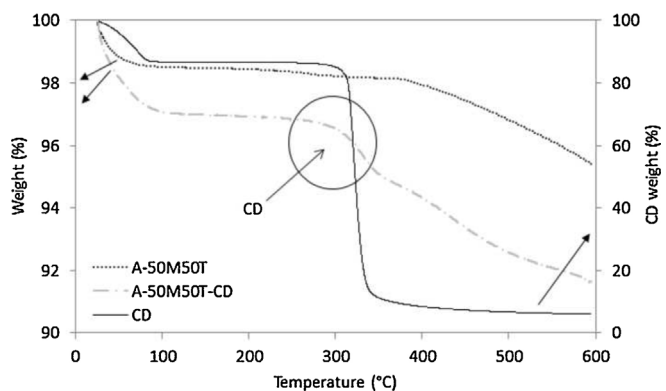


Fig. 2. Comparison of the TGA profiles for the samples A-50M50T, A-50M50T-CD and CD.

angle of these samples, turning them hydrophilic. The weight losses below 100 °C are justified by residual solvent/catalysts evaporation.

The contact angles of the synthesized adsorbents with water are given in Table 2. When the percentage of MTMS in the precursor system increases from 20 to 80% (decreasing the percentage of TMOS in the opposite sense), the contact angles of the aerogels vary from 29 to 125°, *i.e.* the material behaviour spans from hydrophilic to hydrophobic. The xerogel shows a slightly higher contact angle when compared to the equivalent aerogel. This may be explained by a different roughness of the xerogel surface originated by a denser porous network, as will be seen in the next section. The addition of  $\beta$ -CD induces a decrease in the contact angle, which can be explained by the additional OH groups of the macromolecule and, in addition, a possible incomplete condensation of silanols due to the presence of  $\beta$ -CD.

### 3.1.2. Physical and structural characteristics of the adsorbents

Table 2 presents the bulk densities of the obtained aerogels/xerogels, as well as their specific surface areas and average pore sizes. Fig. 3 shows their pore size distributions. It can be concluded that the aerogels bulk densities are between 200 and 350 kg m<sup>-3</sup>, which are typical values for aerogels containing orthosilicates [6,12]. The addition of  $\beta$ -CD does not change significantly the bulk density. For the xerogel, the bulk density is higher (~500 kg m<sup>-3</sup>) and this was expected considering the higher capillary pressures developed during ambient pressure drying, which lead to gel shrinkage.

The specific surface areas of the materials are very high, almost always above 800 m<sup>2</sup> g<sup>-1</sup> (Table 2). In general, a lower bulk

density leads to higher porosity and specific surface area, with the exception of the xerogel for which the higher surface area may be explained by the smaller pore sizes. In fact, it can be seen in Fig. 3 that the pore size distribution of the xerogel shifts to lower pore diameter (30–50 Å), when compared to aerogels, and this can be explained again by the pore shrinkage during evaporative drying. In the case of aerogels, there are two regions of pore sizes with higher frequency: one between 60 and 110 Å, due to the contribution of TMOS in the network, and the other between 100 and 700 Å, explained by the contribution of MTMS (larger pores due to the steric hindrance resultant from the presence of the methyl groups). For all samples, the pore sizes are mainly in the mesopores range (20–500 Å). As for density, the addition of  $\beta$ -CD does not seem to modify significantly the pore size distribution.

SEM micrographs of the obtained materials are presented in Fig. 4. They show very similar and more closed structures for the samples with a percentage of TMOS up to 50%, with samples A-20M80T and X-50M50T showing the densest appearance. Sample A-80M20T clearly has more large pores, in agreement with the pore size distribution results. All the adsorbents are formed by a 3-D solid network with very small interlinked units (<1  $\mu$ m).

### 3.2. Adsorption equilibrium and kinetics of SDBS on the X-50M50T adsorbent

Fig. 5 shows the effect of the initial concentration of SDBS on its adsorption kinetics on X-50M50T. In a first approach, it can be seen that both the amount of SDBS adsorbed by the xerogel and the required time to reach equilibrium increase by increasing the SDBS concentration. The later ranges from 60 to 120 min, when the concentration of SDBS changes from 11.0 to 87 mg L<sup>-1</sup>. It is known that achieving adsorption equilibrium in a short period of time suggests a physical adsorption mechanism, whilst a long range time suggests a chemical-based adsorption process or difficulty of accessing to adsorbent active sites by the adsorbate [34]. Fig. 6 shows representative plots of the fit of linearized forms of Eqs. (3) and (4) to experimental kinetics data. From this analysis, and from the analysis of fitting parameters resumed in Table 3, it can be seen that the pseudo-second order model shows better correlation coefficients. Both facts suggest that adsorption of SDBS by xerogel follows the pseudo-second order kinetic model, which relies on the assumption that a monolayer formation may be the rate-limiting step [35,36], in agreement with the Langmuir sorption isotherms, as will be discussed below.

Rate constants,  $k_2$ , are of same order of magnitude than those found for similar systems [34] and the observed decrease by

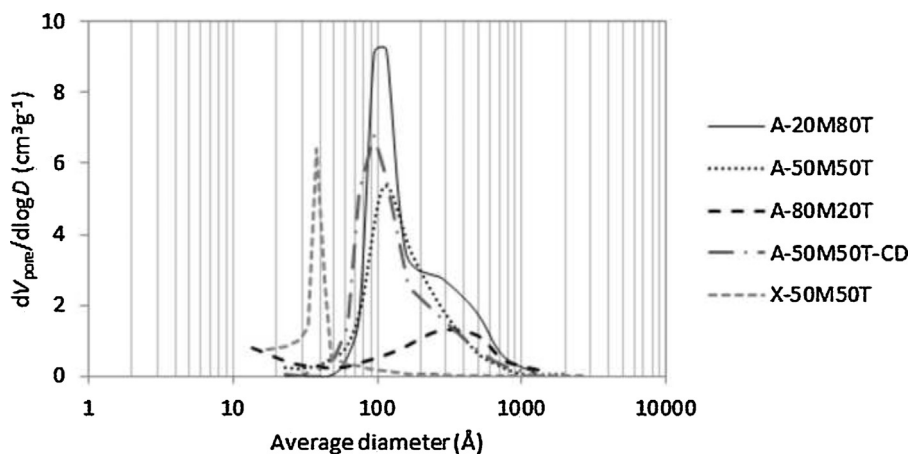
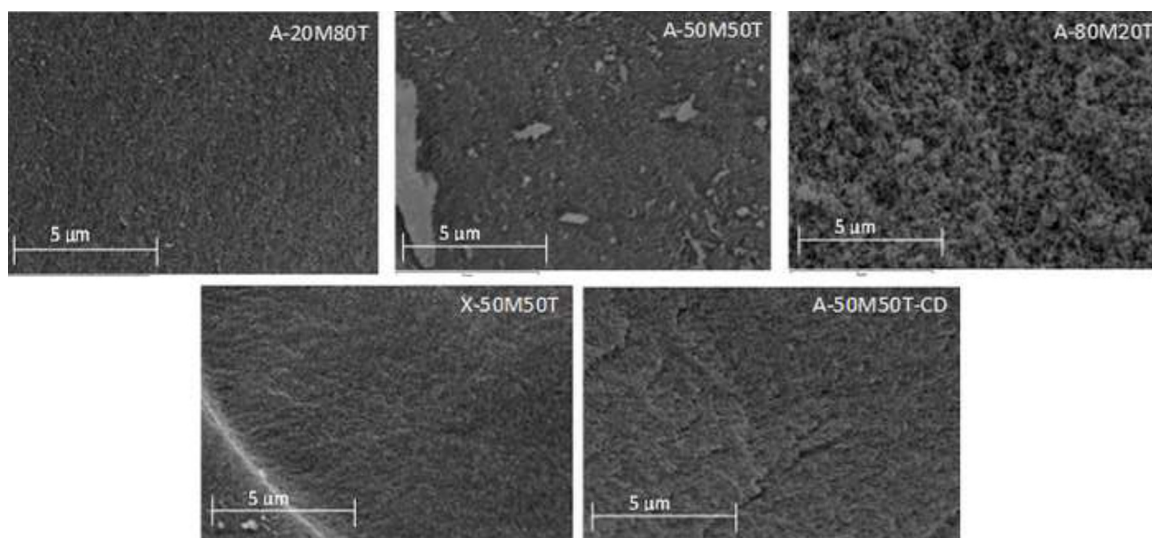


Fig. 3. Pore size distributions of the synthesized adsorbents.

**Table 2**  
Bulk density, contact angle, specific surface area and average pore size of the adsorbents.

Sample	Bulk density ( $\text{kg m}^{-3}$ )	Contact angle ( $^\circ$ )	Specific surface area (BET) ( $\text{m}^2 \text{g}^{-1}$ )	Pore size (BJH desorption) ( $\text{\AA}$ )
A-20M80T	239	$35 \pm 6$	$843 \pm 3$	115
A-50M50T	243	$97 \pm 4$	$888 \pm 3$	111
A-80M20T	351	$120 \pm 5$	$655 \pm 2$	275
A-50M50T-CD	297	$61 \pm 13$	$826 \pm 3$	94
X-50M50T	523	$107 \pm 4$	$938 \pm 3$	38



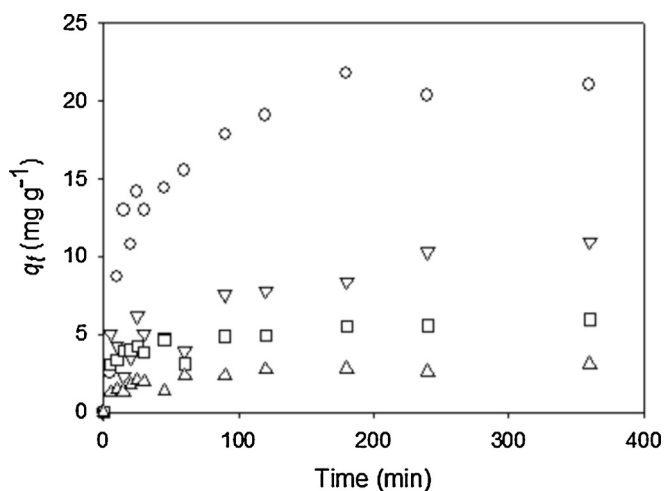
**Fig. 4.** SEM micrographs of the synthesized adsorbents.

**Table 3**  
Kinetic parameters for the adsorption of SDBS on X-50M50T.

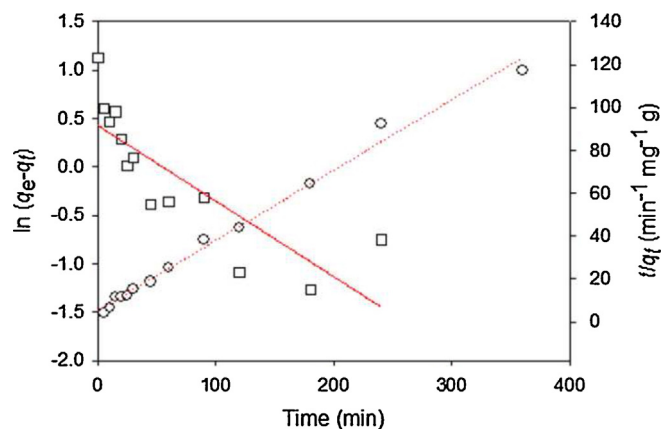
$C_0$ ( $\text{mg L}^{-1}$ )	Pseudo-first order equation			Pseudo-second order equation			
	$k_1$ ( $10^{-3} \text{ min}^{-1}$ )	$q_e$ ( $\text{mg g}^{-1}$ )	$R^2$	$k_2$ ( $10^{-2} \text{ min g mg}^{-1}$ )	$q_e$ ( $\text{mg g}^{-1}$ )	$R^2$	$q_e$ (exp) ( $\text{mg g}^{-1}$ )
11	$8 \pm 2$	$1.5 \pm 0.2$	0.65228	$2.1 \pm 0.1$	$3.05 \pm 0.08$	0.99013	3.1
22	$9 \pm 1$	$2.7 \pm 0.3$	0.81092	$1.4 \pm 0.7$	$6.0 \pm 0.1$	0.99576	$5.7 \pm 0.2$
44	$9.9 \pm 0.9$	$9.5 \pm 0.9$	0.92755	$0.17 \pm 0.06$	$12.1 \pm 0.5$	0.98524	$10.6 \pm 0.5$
87	$17 \pm 1$	$16 \pm 1$	0.90327	$0.21 \pm 0.08$	$22.5 \pm 0.5$	0.99391	$21.1 \pm 0.7$

increasing the initial concentration of SDBS is in agreement with the dependence of time needed to reach the adsorption equilibrium. It is also worth noticing that by increasing the initial concentration of SDBS to 44 and 87  $\text{mg L}^{-1}$ ,  $k_2$  drops one order of magnitude.

In order to have a further insight on the mechanism of SDBS adsorption by X-50M50T, adsorption isotherms were determined and these are shown in Fig. 7. The Langmuir model equation (Eq. (5)) fits well the experimental data ( $R^2 = 0.999$ ), and the following



**Fig. 5.** Adsorption kinetics of SDBS, at different concentrations, on X-50M50T, at 20 °C. ( $\Delta$ ) [SDBS] = 11  $\text{mg L}^{-1}$ , ( $\square$ ) [SDBS] = 22  $\text{mg L}^{-1}$ , ( $\nabla$ ) [SDBS] = 44  $\text{mg L}^{-1}$ , ( $\circ$ ) [SDBS] = 87  $\text{mg L}^{-1}$ .



**Fig. 6.** Representative plot of the fit of linearized forms of pseudo-first (solid line) and pseudo-second (dashed line) order equations to experimental adsorbed amounts of SDBS (with an initial concentration equal to 11  $\text{mg L}^{-1}$ ) by X-50M50T, at 20 °C.

**Table 4**  
Parameters of Langmuir and Freundlich isotherms and maximum percentage of adsorbate removed.

Adsorbate	Adsorbent	Freundlich model			$C_0$ (mg L <sup>-1</sup> )	% Removal
		$K_F$ (mg g <sup>-1</sup> (L mg <sup>-1</sup> ) <sup>1/n</sup> )	1/n	$R^2$		
Phenol	X-50M50T	$2.72 \times 10^{-2}$	1.09	0.993	100	11.2
	A-20M80T	$1.17 \times 10^{-10}$	3.87	0.988	1000	5.7
	A-50M50T	$6.47 \times 10^{-7}$	2.74	0.988	1000	13.2
	A-80M20T	$3.06 \times 10^{-2}$	1.08	0.989	50	11.1
	A-50M50T-CD	$1.16 \times 10^{-4}$	1.89	0.995	1000	8.8
<i>p</i> -Cresol	X-50M50T	$4.30 \times 10^{-3}$	1.42	0.938	10	22.1
	A-20M80T	$1.29 \times 10^{-2}$	1.02	0.921	300	4.6
	A-50M50T	$2.81 \times 10^{-2}$	1.16	0.992	1000	14.3
	A-80M20T	$1.94 \times 10^{-1}$	0.88	0.998	200	18.3
	A-50M50T-CD	$1.84 \times 10^{-1}$	0.68	0.886	50	7.8
4-Chloro-phenol	X-50M50T	$5.24 \times 10^{-2}$	1.07	0.977	10	66.4
	A-20M80T	$7.69 \times 10^{-1}$	0.43	0.751	50	15.5
	A-50M50T	$1.36 \times 10^1$	0.32	0.919	10	65.7
	A-80M20T	$30.0 \times 10^{-1}$	0.48	0.989	50	46.4
	A-50M50T-CD	$26.0 \times 10^{-1}$	0.68	0.986	200	58.5

Adsorbate	Adsorbent	Langmuir model		
		$K_L$ (L mg <sup>-1</sup> )	$q_{max}$ (mg g <sup>-1</sup> )	$R^2$
4-Chloro-phenol	A-50M50T	$1.44 \times 10^{-2}$	117.8	0.985

fitting parameters were obtained:  $K = 4.63 \times 10^{-4}$  L mg<sup>-1</sup> and  $q_{max} = 1029$  mg g<sup>-1</sup>. The empirical Freundlich equation does not fit so well to experimental data ( $R^2 = 0.992$ ), leading to  $K_F = 2.108$  (mg g<sup>-1</sup> (L mg<sup>-1</sup>)<sup>1/n</sup>) and  $1/n = 0.716$ ; a  $n$  value just slightly higher than 1 also confirms that the adsorption of SDBS on the xerogel is modest.

The free energy of adsorption,  $\Delta G_{ads}^0$ , of SDBS onto the xerogel can be computed from the equation:

$$\Delta G_{ads}^0 = -RT \ln(K_0) \quad (7)$$

where  $R$  is the gas constant,  $T$  (=293.15 K) is the absolute temperature and  $K_0 = 1/K$  [37], and is equal to  $-18.7$  kJ/mol. This value suggests that hydrophobic interactions, between the surfactant and the xerogel, play a major role on the interaction mechanism. The analysis of fitting parameters of Langmuir equation also shows that the xerogel has a significant maximum capacity of SDBS adsorption.

Looking to the dependence of the percentage of SDBS removal on the initial concentration of SDBS, we can observe a reduction of the removal percentage from 57 to 43% when the concentration of SDBS increases from 11 to 348 mg L<sup>-1</sup> (pre-micelle concentrations), respectively. Then, the SDBS removal percentage level off

for concentrations above the CMC (e.g., for  $C_0 = 1742$  mg L<sup>-1</sup> the removal percentage is equal to 40.4%). By comparing the SDBS free energy of micellization ( $-32.2$  kJ/mol [38]) with  $\Delta G_{ads}^0$ , it can be concluded that unimer–unimer interaction is stronger than unimer–xerogel and, consequently, adsorption has no effect on the micellization process.

### 3.3. Adsorption of phenolic contaminants into aerogels/xerogel

The adsorption equilibrium data of four aerogels (A-20M80T, A-50M50T, A-80M20T and A-50M50T-CD) and one xerogel (X-50M50T) for the selected toxic phenolic compounds are shown in Figs. 8–10. From global results, it can be seen that the tested materials exhibit different adsorption capacities for phenol, *p*-cresol and 4-chlorophenol depending on their level of hydrophobicity determined by amounts of MTMS and TMOS precursors incorporated in the synthesis process.

The unfavourable nature of most adsorption isotherms obtained for phenol and *p*-cresol, as illustrated in Figs. 8 and 9, suggests limited affinity between these contaminants and the studied

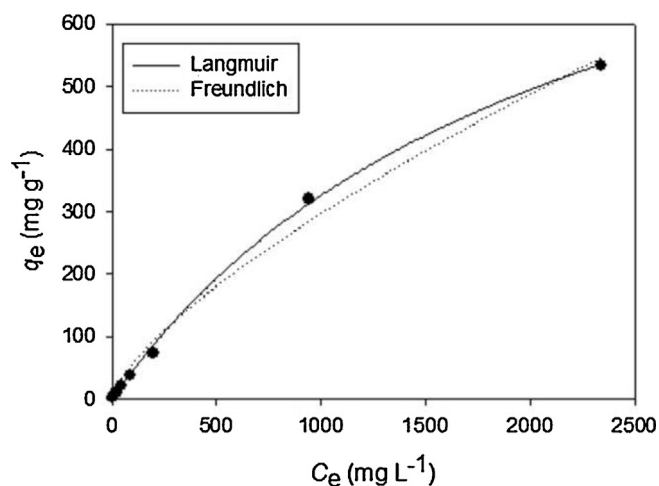


Fig. 7. Adsorption isotherms for X-50M50T/SDBS system, at 20 °C.

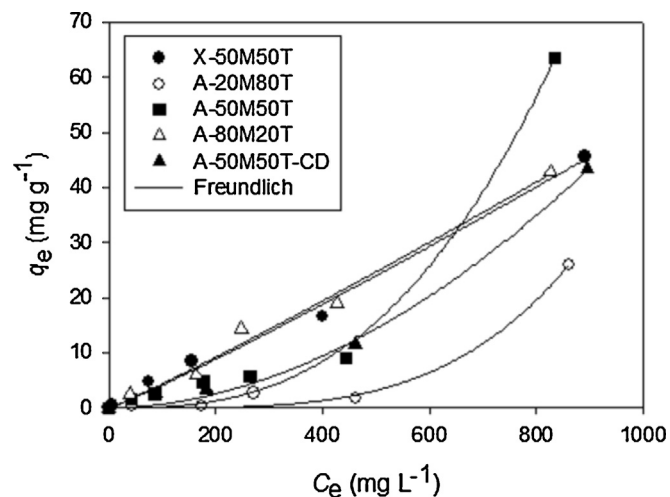


Fig. 8. Experimental and calculated equilibrium data for adsorption of phenol on aerogels and xerogel adsorbents.



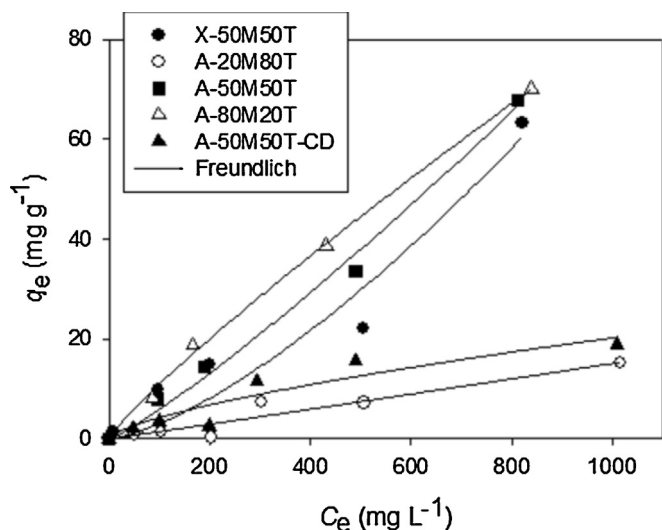


Fig. 9. Experimental and calculated equilibrium data for adsorption of *p*-cresol on aerogels and xerogel adsorbents.

aerogels/xerogel, mainly for low concentrations. Fig. 10 shows that 4-chlorophenol is adsorbed to a greater extent than the other compounds, and the equilibrium data is characterized by favourable isotherms. This should be attributed to the stronger hydrophobic character of the adsorbate that plays an important role in the amount adsorbed from aqueous solutions. The modification of the aerogel with  $\beta$ -CD (A-50M50T-CD) was successful in the case of 4-chlorophenol (see Fig. 10) likely because its hydrophobic interactions with the  $\beta$ -CD cavities in the adsorbent were enhanced. Moreover, it is also worth mentioning that higher adsorption capacity is observed for the aerogel-type adsorbents than for the xerogel, for all the adsorbates. The higher amount of mesopores in the aerogels when compared to the xerogel seems to play an important role in the adsorption process. Effectively, the very fine porous structure of the xerogel (Section 3.1.2) may hinder the diffusion of the phenolic compounds in the solid matrix.

The parameter values estimated by fitting the equilibrium models to the experimental data are listed in Table 4. It should be noted that the Langmuir model only fits well the data corresponding to the system A-50M50T/4-chlorophenol. Therefore, the Freundlich model is the most suitable, which has been applied in numerous cases involving the removal of organic contaminants from aqueous solutions since it has the ability to describe the adsorption in heterogeneous surfaces. The correlation coefficients ( $R^2$ ) shown in

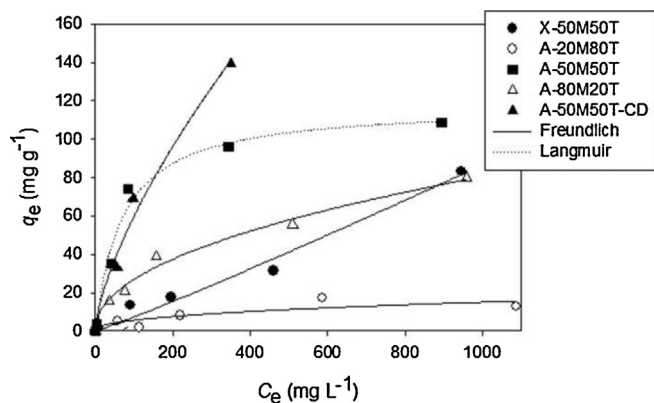


Fig. 10. Experimental and calculated equilibrium data for adsorption of 4-chlorophenol on aerogels and xerogel adsorbents.

the table indicate that the experimental and predicted results are in good agreement in most cases.

With respect to the maximum percentage of adsorbate removed from the aqueous solution, Table 4 shows the initial concentrations,  $C_0$ , for which the best results were observed for each pollutant. In the case of phenol, the percentage of removal was low for all the tested materials, but the adsorbent A-50M50T revealed to have the best performance. Contrarily to our expectations, the addition of  $\beta$ -CD does not improve the performance of the aerogel A-50M50T. Nevertheless, the percentages of phenol removed in our investigation are higher than in other studies reported in the literature [8,16], where for a solution with initial concentration of  $1000 \text{ mg L}^{-1}$ , the equilibrium adsorption was around  $10 \text{ mg g}^{-1}$  and the corresponding percentage of removal was close to 4%. Regarding *p*-cresol, the percentage of removal improves slightly for A-50M50T when compared to phenol. However, also for this compound further improvement in the adsorbents will be required. Finally, the best results were obtained for 4-chlorophenol, since removals higher than 60% were observed. As mentioned before, it should be noted that the addition of  $\beta$ -CD seems to be an interesting route to improve the removal of this phenolic compound.

#### 4. Conclusions

The silica-based aerogels and xerogels prepared in this work exhibit mesoporous structures with high surface area and tailored levels of hydrophobicity/hydrophilicity. This last feature is achieved by combining different amounts of methyltrimethoxysilane and tetramethylorthosilicate co-precursors, as well as by functionalizing the silica aerogels with  $\beta$ -cyclodextrin macrocycle.

The xerogel with intermediate wetting behaviour shows good removal efficiency for SDBS and a pseudo-second order adsorption kinetics. In this case the adsorption is probably characterized by hydrophobic interactions and the monolayer formation may be the rate-limiting step. When studying the aerogels as adsorbents for several phenolic compounds – phenol, *p*-cresol and 4-chlorophenol, the hydrophobic adsorbents show higher performances for the adsorbates with higher partition coefficient octanol/water, with favourable isotherms, showing that the hydrophobic interactions play an important role in the adsorption process. In particular, the aerogel modified with  $\beta$ -cyclodextrin leads to high adsorption capacity of 4-chlorophenol, the most hydrophobic adsorbate tested, being this a result of the high hydrophobic character of the  $\beta$ -cyclodextrin cavity.

#### Acknowledgements

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