

1 **Advances and challenges in retinoid delivery systems in regenerative and therapeutic**
2 **medicine**

3
4 Running title: RA delivery systems

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37 **Highlights**

38

39 • Recent discoveries of **retinoid** action in cancer, stem cell and immune cell biology have been
40 translated into a high number of clinical trials

41 • Controlled release systems are important for clinical translation of **retinoids** because they
42 increase its aqueous solubility and lifetime in the bloodstream, while decreasing RA photo-
43 sensitiveness, cytotoxicity and side effects

44 • The properties of **retinoid**-controlled release systems (i.e. physicochemical, release, targeting
45 and uptake/intracellular trafficking properties) vary according to their ultimate biomedical
46 applications

47 • Further developments in **retinoid** controlled-release systems are needed to increase
48 organ/tissue/cell targeting, as well as capacity to release **retinoids** in combination with other
49 drugs

50 **Abstract**

51 **Retinoids** regulate a wide spectrum of cellular functions from the embryo throughout
52 adulthood, including cell differentiation, metabolic regulation and inflammation. The development
53 of **retinoid** delivery systems offers several advantages for clinical translation of **retinoid**-based
54 therapies, including improved solubilization, prolonged circulation, reduced toxicity, sustained
55 release and improved efficacy. In this Review, we discuss advances in preclinical and clinical tests
56 regarding **retinoid** formulations, **specifically the ones based in natural retinoids**, evaluated in the
57 context of regenerative medicine, brain, cancer, skin and immune diseases. Advantages and
58 limitations of **retinoid** formulations as well as future prospects will be presented.

59

60 **Keywords**

61 **Retinoid**, retinoic acid, delivery systems, cancer, inflammation, brain pathologies, skin diseases

62

63

64 **1. Introduction**

65 Retinoic acid (RA) signaling is one of the most important biological pathways in nature,
66 triggered by RA interaction with nuclear receptors that control gene expression. The chemical
67 structure of retinol (vitamin A, a RA precursor) was first described by Paul Karrer in 1931¹, who
68 was awarded a Nobel prize in 1937 for the discovery. The use of RA for skin disorders² and cancer
69 treatment (acute myeloid leukemia³ and cervical neoplasia^{4,5}) started in the 1960's and 1980's
70 (**Figure 1**). In 2000's, RA had been incorporated in many tissue engineering scaffolds as a stem cell
71 differentiation agent^{6,7}. Over the last 10 years, many discoveries related to the biological role of RA
72 in controlling the biology of hematopoietic stem cells^{8,9}, tumor-initiating cells¹⁰⁻¹², immune cells¹³,
73 intestinal mucosa wound repair¹⁴, cancer resistance¹⁵ and cell reprogramming and
74 differentiation^{16,17}, have further stimulated interest in this drug for many other biomedical
75 applications. This interest is confirmed by more than 50 active clinical trials (according to
76 ClinicalTrials.gov) evaluating the effect of RA in cancer (28 trials), mostly in hematological (16
77 trials) and brain tumors (8 trials), skin pathologies (e.g. acne, photoaging, eczema) (5 trials), as well
78 as in other conditions such as inflammation, olfactory loss and neuropsychiatric diseases (**Table 1**).

79 Clinical applications of RA have highlighted three main limitations of its pharmacological
80 use. First, RA is poorly soluble in aqueous solutions¹⁸ and photosensitive¹⁹, which makes its
81 administration challenging. Secondly, RA induces irritation when applied onto skin and increases
82 its catabolism, when it is administered intravenously, reducing its therapeutic efficacy²⁰. Lastly, RA
83 is involved in many biological processes and thus the systemic delivery of RA causes side effects.
84 All these limitations motivated researchers to **synthesize novel and better tolerated synthetic**
85 **retinoid compounds** and to develop **retinoid** delivery formulations based on gels, liposomes,
86 microparticles, nanoparticles, and micro-/nanofibers **which, in some cases, have been** modified to
87 target specific tissues and cells of interest²¹.

88 Recent developments in the use of **retinoid** for cancer treatment^{22,23} and differentiation
89 studies using stem cells^{8,24}, as well as in the development of more advanced formulations to control
90 its bioactivity^{25,26} makes this review timely. In addition, with the exception of a limited number of
91 reviews, with a restricted scientific scope in **retinoid** formulations²⁷ or highlighting the importance
92 of **retinoids** in development²⁸ or for specific therapeutic applications²⁹, no study has fully covered
93 the application of **retinoid** formulations for therapeutic and regenerative medicine applications. In
94 this review, the role of **retinoid** formulations in the context of the brain, skin, immune system and
95 cancer applications will be discussed. For each application, the pathological context will be briefly
96 presented as well as the effect of **retinoid** administered without any controlled release system. The
97 reasons behind the development of **retinoid** formulations in each application will be presented, as

98 well as their benefits and limitations, considering retinoid solubility, photostability,
99 biocompatibility, release profile, tissue/cell availability and targeting, and therapeutic efficacy.

100

101 2. Retinoid chemistry and general overview on RA signaling

102 2.1. Retinoid chemistry

103 Retinoids are a class of compounds composed by 3 regions: a hydrophobic, a central
104 polyene and a polar (usually a carboxyl group)²⁹. There are 3 natural retinoids: all-*trans* RA
105 (ATRA), 9-*cis* RA (alitretinoin) and 13-*cis* RA (isotretinoin). To decrease the toxicity of the natural
106 retinoids as well as to increase their stability and selectivity against a specific RAR subtype, several
107 semi-synthetic and synthetic retinoids (including “atypical retinoids”) have been developed^{29,31}.
108 Some of them have been tested in clinical trials and approved for therapy (Table 2). At least 4
109 molecules have reached phase 4 clinical trials: bexarotene, tazarotene, adapalene and
110 trifarotene^{29,31,32}. Bexarotene has optimal RXR binding, which effectively causes cancer cell death,
111 particularly in cutaneous T cell lymphoma. However, high levels of serum aminotransferase and
112 liver injury have been reported³³. Tazarotene and adapalene formulations have high affinity and
113 selectivity for RAR β and RAR γ , although the first one showed more toxicity than the second in
114 dermatological applications³⁴. Trifarotene is a selective RAR- γ agonist, approved in late 2019 in the
115 USA, for the topical treatment of acne in patients as young as 9 years old³². Despite the progresses
116 made in the last years in the synthesis of novel retinoids to increase natural retinoid efficacy while
117 reducing their toxicity, it is evident that natural retinoids are still under intense scrutiny as
118 demonstrated by a considerable number of clinical trials (Table 1). It is likely that the reasons are
119 combinatorial: RA is a natural drug, blocks multiple disease-signaling pathways¹², in opposition to
120 some synthetic retinoids that are very selective to a single receptor-mediating signaling pathway,
121 and it has been used for many years in combinatorial therapy and thus well known by clinicians.
122 Because it would be difficult to cover all advances made in the area of retinoid delivery systems in a
123 single review, the authors have chosen to cover only natural retinoids in the present manuscript.
124 This focused approach is also supported by recent progresses in RA biology and many formulations
125 developed in the last years for the delivery of RA.

126

127 2.2. RA signaling

128 RA is the main biologically active metabolite of vitamin A²⁸. In humans, the only source of
129 vitamin A is obtained through diet, as lipophilic retinol (or its more stable form, retinyl ester) or as
130 carotenoids. The transport of these retinoids to cells occurs when blood circulating retinol is bound
131 to retinol-binding protein (RBP) 4 (Figure 2). This complex interacts with membrane transporter

132 and receptor *stimulated by retinoic acid 6* (STRA6) facilitating entry into the cytoplasm, where
133 retinol binds to Crbp1 (encoded by RBP1). A two-step process converts retinol into ATRA. RA
134 binds to cellular retinoic acid binding-proteins (CRABP) , assisting autocrine and paracrine
135 signaling³⁵. The mechanisms underlying paracrine signaling remain unclear while autocrine
136 signaling requires CRABP2 for nuclear entry^{28,35}. In the nucleus, RA triggers gene transcription by
137 binding to **heterodimers formed by** RA receptors (RAR α , RAR β and RAR γ) **and retinoid X**
138 **receptors (RXR α , RXR β , and RXR γ).** **RAR-RXR heterodimers interact with** a deoxyribonucleic
139 acid sequence known as the retinoic acid-response element (RARE)³⁶, **which facilitates the binding**
140 **of co-activators with histone acetylase activity, ultimately leading to the transcription of target**
141 **genes (Figure 2).** Recent data showed that RXR α homodimers can also regulate gene
142 **transcription³⁷.** Another isomer of RA, isomer 9-*cis* RA (alitretinoin), binds to retinoid X
143 receptors^{28,36}, while 13-*cis* RA (isotretinoin), has negligible affinity for retinoic acid receptors (RAR
144 or RXR) or for cellular RBP³⁸. However, 13-*cis* RA may be converted into molecules that act as
145 agonists for nuclear RAR and RXR. **Importantly, RAR also regulates non-nuclear and non-**
146 **transcriptional effects, namely the activation of kinase signaling pathways³⁹ (Figure 2).** Finally, RA
147 is catabolized by monooxygenases of the cytochrome P450 superfamily³⁵.

148

149 **3. Formulations for the delivery of RA**

150 RA has very low solubility in aqueous solution (0.21 μ M at pH 7.3)¹⁸ and thus requires
151 specific binding proteins (e.g. CRABPs) to be transported within cells to act at nuclear receptors. In
152 addition, it has a short (few hours) lifetime, due to its degradation by a cytochrome P450-dependent
153 monooxygenase system⁴⁰. Moreover, it induces undesirable side effects (congenital malformations⁴¹
154 as well as mucocutaneous dryness, headache, and hypertriglyceridemia⁴²) when administered at
155 high concentrations. Therefore, for more than 35 years, several groups have developed different RA
156 delivery systems to overcome these limitations. Delivery systems based in polymeric scaffolds (e.g.
157 hydrogels, nanofibers), nanoparticles (liposomes, micelles, polymeric, dendrimers), microparticles,
158 among others, are described and summarized in **Table 3**.

159

160 **3.1. Type and strategies for the delivery of RA**

161 Several strategies have been used to prepare RA delivery systems: namely, by complexation
162 of RA with proteins (e.g. transthyretin)⁴³ or cationic polymers (e.g. poly(ethyleneimine)(PEI))^{44,45},
163 by physical encapsulation in polymeric^{25,45} or inorganic⁴⁶ nanoparticles, microparticles⁴⁷,
164 micelles^{48,49}, liposomes⁵⁰ or films⁴⁴, by covalent attachment of RA to a carrier⁵¹, or by
165 immobilization of RA on surfaces of nanoparticles⁴⁶, among others. Despite several approaches

166 having been reported of RA delivery systems based on polymeric scaffolds or electrospun fibrous
167 meshes, the most common strategy for RA delivery has been based on liposomal or polymeric
168 nanoparticles formed by polyesters, polyimines, polysaccharides and proteins (Table 3). A
169 significant number of formulations have allowed the RA encapsulation in the core of the
170 nanoparticles. This RA encapsulation was either obtained by (i) physical or (ii) chemical interaction
171 with the components of the nanoparticle or (iii) by physical entrapment. Regarding physical
172 interaction, RA is complexed with positively charged polymers such as chitosan, or PEI^{44,52}. The
173 carboxylic acid of RA interacts electrostatically with the amine group located in the polymer,
174 forming a complex that can be stabilized by addition of a polyanion and divalent ions^{44,45}.
175 Concerning chemical interaction, RA is typically conjugated chemically to one of the components
176 of the nanoparticle by biodegradable ester^{51,53}, amide^{26,54-56} or disulfide⁵⁷ linkages. These bonds are
177 susceptible to degradation during specific pH conditions, in presence of proteases, or reducing
178 agents leading to the RA release. Concerning physical entrapment, RA is captured during
179 nanoparticle formation^{58,59} or in the pores of the nanoparticles⁶⁰. In a relatively low number of
180 formulations, RA was immobilized not in the core but on the surface of the nanoparticle²²; however,
181 the immobilized concentration of RA is lower than the ones having RA in the core. Formulations
182 with high (above 100 µg/mg of formulation)^{52,57,61}, medium (between 100 and 25 µg/mg of
183 formulation)^{45,46} and low (below 25 µg/mg of formulation)^{7,59} loading have been described.

184 The release profile of RA depends on several factors of the formulation, including the size,
185 composition, initial concentration of RA loaded and its degradation profile. Studies have reported
186 the sustained release of RA for more than 1 month by encapsulating it into biodegradable
187 microspheres and tuning the release rate by adjusting polymer composition in the formulation⁶², by
188 the encapsulation in polyion complex micelles⁶³ and adjusting polymer composition or drug
189 content, or by the encapsulation in liposomes⁵⁰. In general, formulations with high RA loading
190 showed a slower release profile of the drug⁵². Because of the hydrophobicity of RA, the occurrence
191 of a burst release in most formulations is negligible^{45,48,52,64}.

192 Recent developments to improve the delivery of RA have led to the design of formulations
193 that can be controlled remotely (temporally and spatially) by an external stimulus such as light,
194 ultrasound or magnetic forces⁶⁵. These stimuli-responsive biomaterials are suitable to control RA
195 kinetics delivery. In that sense, light-activatable nanoparticles containing RA that disassemble in
196 minutes after activation by a blue laser at 405 nm have been prepared^{25,26,61}. Importantly, the
197 formulations presented higher activity than formulations in which RA was released by passive
198 diffusion (not light-triggered), because they rapidly saturated nuclear receptors.

199

3.2- Capacity of RA-containing formulations to overcome biological barriers

The cellular uptake of some RA-containing nanoparticles has been demonstrated to take below 12 h⁴⁵ by clathrin-mediated endocytosis and macropinocytosis²⁵. Formulations that escaped the endolysosomal compartment accumulated in the cytoplasm in less than 24 hours^{25,46}. It is possible that RA released in the cell cytoplasm binds to transport proteins such as cellular retinoid-binding protein II (CRABP-II) and/or fatty acid-binding protein 5 (FABP5), followed by its transportation to the cell nucleus⁶⁶. The intracellular concentration of formulations containing RA was dependent on the initial formulation loading, type of formulation, and type of cell^{25,46}. Uptake between 25 and 80 pg of nanoparticles containing RA *per cell* has been described²⁵.

The capacity of RA-containing formulations to cross biological barriers such as the blood-brain barrier (BBB) (relevance for the treatment of brain cancer and neurodegenerative disorders) is a topic largely unexplored. In most cases, the formulations have been administered by stereotaxic and not intravenous administration⁶⁷. Experimental data in humans indicate that ATRA administered orally is not able to accumulate in the cerebrospinal fluid⁶⁸. Clinical trials such as NCT00528437 and others are now investigating the pharmacokinetics of 13-*cis* RA and its accumulation in the cerebrospinal fluid.

3.3. Pre-clinical and clinical applications of RA-containing formulations

Formulations containing RA have been used both in pre-clinical and clinical trials (**Figure 3**). Most pre-clinical tests were performed in mice in the context of regenerative medicine^{46,61,67,69} and cancer applications^{22,25,57}. *In vitro* tests showed that ATRA-containing liposomes⁷⁰ or ATRA-containing polymeric nanoparticles^{25,71} were 100-1000 times more active than soluble ATRA in cultured tumor cells^{25,70} or neural stem cells⁷¹. *In vivo* tests showed that rats⁵⁰ or human patients⁷² treated with ATRA-containing liposomes by intravenous administration had no decrease in plasma ATRA levels while animals/humans treated with oral formulation of ATRA (non-liposomal) had a significant decrease in plasma ATRA levels^{42,50}. The results indicate that the hepatic metabolism of ATRA encapsulated in liposomes was inferior to the one observed in ATRA administered orally. Both formulations were safe in human trials. The clinical application of RA-containing formulations can be divided into 2 groups: (i) topical and (ii) oral administration. For topical administration, seven RA-containing formulations have reached the market for the treatment of skin-related diseases (**Table 4**). Here, current progresses are concentrated in reducing the toxicity of RA (e.g. by the use of synthetic retinoids³²), and exploring the combination of RA with other drugs (see **Table 4**). For oral administration, there are four formulations containing RA that have reached the market, particularly for the treatment of skin diseases (three of them) and cancer (only one)

234 (Table 4). Others were tested in clinical trials but did not reach the market. For example, a
235 liposomal-based formulation of RA (monotherapy) was successfully tested in patients with facial
236 acne⁷³ and refractory hematological malignancies in a phase II clinical trial⁷². In the last application,
237 the remission results (67% clinical remission) were lower than a combinatorial therapy (77%
238 clinical remission) based on oral administration of ATRA plus idarubicin and thus the formulation
239 was not further evaluated. Unfortunately, the liposomal formulation of RA was not tested in
240 combination with other drugs, as it is now being investigated with non-liposomal RA formulations
241 (Table 1), and this deserves further investigation in the near future.

242

243 4. RA delivery systems for Regenerative Medicine

244 4.1. Embryonic stem cells

245 One of the initial applications of RA for Regenerative Medicine was as a differentiation
246 agent during embryogenesis. The spatiotemporal release of RA by polymeric microparticles
247 incorporated within embryonic bodies, derived from human embryonic stem cells, was reported to
248 induce cell differentiation and tissue formation resembling the phenotype and structure of early
249 human embryos⁷. Initial studies have used RA as a potent regulator of neural differentiation⁷⁴. RA
250 downregulates expressions of geminin and zinc finger protein Zic2, SoxB1 (Sox-1, Sox-2, Sox-3)
251 and Notch-1, which maintain neural progenitor cell proliferation. By halting proliferation, RA shifts
252 signaling towards differentiation. Several platforms have been used for RA delivery alone or in
253 combination with other agents. For example, RA-containing electrospun fibrous meshes and
254 scaffolds reportedly have an improved effect on stem cell dynamics. Electrospun poly(ϵ -
255 caprolactone) (PCL) fibers were loaded with ATRA, which led to extremely high local
256 concentrations of this agent and to differentiation of murine embryonic cells, within the
257 multilayered scaffolds⁷⁵. Besides the neurogenic potential of RA, recent studies have highlighted
258 the critical role of RA in the derivation of embryonic hematopoietic cells⁹, lymphoid organs¹³ and
259 Langerhans cells that reside specifically in the epidermis¹⁶. Particularly, *ex vivo* activation of RA
260 signaling in hemogenic endothelium, a small subpopulation of endothelial cells that can
261 differentiate into hematopoietic cells, increased its transition into a pool of hematopoietic stem
262 cells. Conversely, RA pathway shutdown terminated this process⁹. Additionally, fetal type 3 innate
263 lymphoid cells (ILC) are modulated by RA signaling *in utero* and therefore depend on appropriate
264 maternal dietary intake of retinoids throughout pregnancy. This period also determines the ability of
265 ILC progenitors to differentiate into mature lymphoid tissue-inducing cells¹³. It is possible that
266 some of the RA delivery systems developed so far may be used for embryonic immune and
267 hematopoietic stem cell development studies.

268

269 **4.2. Adult stem cells**

270 RA is also an important regulator of adult stem cells. For example, ATRA antagonizes
271 stress-induced activation of dormant hematopoietic stem cells by restricting protein translation and
272 oxidative stress⁸. When mice were fed a vitamin A-free diet to deplete the RA reservoir, animals
273 suffered, among other effects, functional impairment of hematopoietic stem cells and their numbers
274 were unable to recover even after injection with an immunostimulant⁸. In addition, RA-based
275 formulations have been used as inflammatory modulators of stem cells. For example, human
276 mesenchymal stem cells exposed to ATRA-loaded solid lipid nanoparticles significantly reduced
277 IL-6 and IL-8 expression⁷⁶. ATRA-containing nanoparticles have been also developed to deliver
278 RA into neural stem cell (NSC) niches⁴⁵. The nanoparticles had a higher effect on neuronal
279 differentiation than solubilized RA both *in vitro* and *in vivo*^{45,67}. The effect was mediated by an
280 increase in transcription of the pro-neurogenic genes Ngn1 and Mash1^{45,67}. The formulation was
281 then modified to remotely disassemble and release RA with spatial and temporal resolution,
282 triggered by exposure to blue light⁶¹. A single short pulse of light prompted β -catenin-dependent
283 neuronal differentiation and RAR α upregulation. The combined action of blue light and RA
284 enhanced endogenous neurogenesis.

285

286 **5. RA delivery systems for skin diseases**

287 **5.1. RA mode of action**

288 Several RA drugs are available clinically for dermatological treatments, including ATRA
289 and 9-*cis*-RA, among others⁷⁷. For example, 9-*cis*-RA encapsulated in a gel is a FDA-approved
290 topical agent for cutaneous Kaposi's sarcoma⁷⁷. ATRA also has been tested in clinical trials for
291 treatment of the same disease⁷⁸. This sarcoma is associated with human immunodeficiency virus
292 infection and is characterized by a vascular endothelioma (i.e. tumor of the endothelial cells).
293 ATRA is used clinically for treating photoaging, acne and psoriasis. Indeed, ATRA was approved
294 for acne vulgaris treatment in 1971², and since other drugs (called retinoids because they bind to
295 RAR and/or RXR receptors) have been developed. In these clinical applications, the biological
296 effect of ATRA includes: (i) modulation of proliferation and differentiation of skin cells; (ii) anti-
297 inflammatory activity⁷⁷. The existence of several types of receptors and their combinations as
298 heterodimers, as well as ability of ATRA to modulate the activity of multiple kinase signaling
299 pathways independently of the nuclear activation of RAR and RXR receptors, may explain the
300 diversity of ATRA biological actions³⁶. ATRA also induces skin angiogenesis and collagen

301 deposition, increases the mitotic activity of inter- and follicular epithelium, and reduces melanin
302 production⁷⁷.

303

304 **5.2. Type of RA-containing formulations**

305 Conventional formulations containing ATRA require multiple applications to maintain the
306 therapeutic effect. Therefore, several formulations have been developed to improve the long-term
307 effect of ATRA, reducing side effects like desquamation and erythema, skin irritation, and
308 increasing the stability of RA to light¹⁹ (**Figure 4**). In this sense, formulations including
309 phospholipid-based particles (e.g. solid lipid nanoparticles and nanostructured lipidic carriers)^{2,79},
310 polymeric nanoparticles^{2,27,79,80} or polymers conjugated with RA⁵³ have been developed to
311 overcome these issues. The carriers presented a particle size between 100 and 400 nm, a range of
312 zeta potential from neutral (liposomes) to negative (polymeric nanoparticles, ethosomes, solid lipid
313 nanoparticles and nanostructured lipidic carriers), high entrapment efficiency (above 65%), high
314 photostability (between 2 and 3-fold higher than commercial tretinoin dissolved in ethanol),
315 moderate to high skin permeation, high skin tolerance and moderate to high anti-psoriatic activity.
316 Some of these formulations are easier to scale up (e.g. solid lipid nanoparticles) than others^{79,81}. In
317 addition, some formulations (e.g. ATRA-containing liposomes) improve the local effect of RA in
318 the skin and decrease systemic adsorption⁸⁰. Moreover, ATRA-containing formulations increased
319 significantly the chemical stability of ATRA during normal storage conditions (e.g. stability for 1
320 year at 25°C) and after exposure to UV irradiation (2-fold lower photo-degradation)^{79,82}. The
321 protective effect of these formulations was linked to their ability to reflect and scatter UV
322 radiation⁸². Interaction of ATRA with a lipophilic amine (stearylamine) decreases ATRA
323 crystallinity, leading to a formulation with less skin irritating properties⁷⁹.

324

325 **5.3. Pre- and clinical applications**

326 RA-containing formulations are in clinical evaluation for the treatment of *acnes vulgaris*,
327 hand eczema and photoaging (**Tables 1 and 3**). **An ATRA-loaded liposomal formulation has been**
328 **tested in a pilot trial⁷³. Patients with facial acne treated with the liposomal formulation showed**
329 **higher lesion improvements than with conventional formulations. The higher efficacy of the ATRA-**
330 **loaded liposomal formulation was attributed to enhanced penetration of RA across the *stratum***
331 ***corneum*. A commercial tretinoin gel microsphere formulation has reached the market, consisting of**
332 **microspheres with 10-20 μm in size⁷⁷**. A common issue in design of controlled RA release systems
333 for skin applications is limited efficacy in terms of cell targeting (**Figure 4**). Formulations such as

334 micro- and nanoparticles might have the capacity to target specific cells by specificities in their
335 physicochemical properties or by incorporating in their surface peptides, proteins or aptamers that
336 recognize specific cell receptors. Due to their size, these formulations might accumulate
337 preferentially in some regions of the skin, such as the follicular adduct and thus act in those
338 biological environments. Indeed, follicular targeting is important for treatment of acne, because it
339 increases the therapeutic effect of retinoids, while reducing their potential side effects. Polymeric
340 micelles of diblock methoxy-poly(ethylene glycol)-poly(hexyl-substituted lactic acid) copolymer
341 favor follicular targeted delivery of ATRA⁸³.

342

343 **6. RA delivery systems for brain diseases**

344 **Several formulations containing RA have been used to treat Alzheimer's disease (AD),**
345 **Parkinson's disease (PD) and stroke (Table 1).** Current challenges in the use of RA formulations for
346 **brain diseases are ascribed to low accumulation in the brain after intravenous administration**
347 **(Figure 5).** Although the formulations having affinity ligands in their surface have increased
348 **retention in the brain, only a minor fraction of the injected formulation reaches the brain and crosses**
349 **the BBB. Cell-mediated delivery (e.g. T cells, monocytes) may be used to overcome this issue: cells**
350 **can act as transporters of formulations that will be activated by local (e.g. pH) or remote triggers**
351 **(e.g. light, ultrasound, magnetism)^{25,84} once they reach the target site.**

352

353 **6.1. Alzheimer's disease**

354 AD is a neurodegenerative disease caused by accumulation of amyloid-beta peptides,
355 hyperphosphorylated tau filaments and brain vascular changes leading to cerebral amyloid
356 angiopathy⁸⁵. In experimental models of AD, RA: (i) inhibited oxidative damage and mitochondrial
357 dysfunction; (ii) increased ApoE expression and suppressed inflammation; and (iii) improved
358 learning and memory^{86,87}. Nanoparticles have been developed to efficiently deliver ATRA and
359 small interfering RNA to promote NSC differentiation in AD⁸⁸. Transplantation of nanoparticle-
360 treated NSCs in AD mice improved cognition and memory.

361

362 **6.2. Parkinson's disease**

363 **PD is characterized by selective degeneration of dopaminergic neurons in the *substantia***
364 ***nigra* and by accumulation of α -synuclein and Lewy bodies (protein inclusions in neurons)⁶⁹.** In
365 experimental models of PD, RA protected against neurodegeneration of midbrain dopaminergic
366 neurons in the *substantia nigra*^{89,90}. Stereotaxic injection of ATRA-containing nanoparticles in the
367 striatum protected nigral dopaminergic neurons in an *in vivo* PD model⁶⁹. Accordingly, RA-

368 containing nanoparticles increased expression of Nurr1 and Pitx3, key regulators of dopaminergic
369 neuronal development and maintenance^{69,91}.

370

371 **6.3. Stroke**

372 In the case of stroke, changes in the vasculature, or in BBB permeability or function, may
373 cause or enable progression of CNS diseases⁹². RA/RAR signaling is critical for BBB
374 differentiation and integrity⁹³. Recent studies have shown protective effects of ATRA-containing
375 nanoparticles in stroke. The formulation enhanced endothelial cell proliferation and tubule network
376 formation and protected against ischemia-induced death in endothelial cell lines and in endothelial
377 progenitor cells isolated from ischemic stroke patients²⁴. Moreover, when intravenously injected,
378 RA formulations restored neuronal and vascular functions in a prenatal model of brain ischemia⁹⁴.

379

380

381 **7. RA delivery systems for the treatment of cancer**

382 **7.1. Blood cancers**

383 **7.1.1. RA mode of action**

384 One of the first applications of RA delivery systems was for treatment of blood cancers. The
385 antitumor activity of ATRA was demonstrated in 1980 in acute promyelocytic leukemia (a subtype
386 of acute myeloid leukemia (AML) accounting for 5% of AML cases)³. Since then, many clinical
387 trials have evaluated the antitumoral efficacy of the drug alone or in combination with arsenic
388 trioxide or idarubicin⁹⁵. RA promoted terminal differentiation of leukemic cells, while reducing
389 their proliferation. The antitumoral activity of RA (enhanced by cooperation with arsenic trioxide)¹¹
390 was due to inhibition and degradation of prolyl isomerase Pin1, which has a critical role
391 coordinating multiple phosphorylation events during oncogenesis¹². Therefore, RA has the unique
392 property of blocking multiple cancer-driving pathways simultaneously. Unfortunately, the
393 therapeutic efficiency of ATRA-based therapies remains to be demonstrated in patients with AML
394 without acute promyelocytic leukemia.

395

396 **7.1.2. Type of RA-containing formulations**

397 Physicochemical properties as well as release properties of RA-containing formulations for
398 treatment of blood cancers are summarized in **Table 3**. With exception of some formulations for
399 parenteral administration⁶⁴ and for intracellular delivery in *ex vivo* conditions^{25,26}, most RA-
400 containing formulations have been developed for intravenous delivery. These formulations were

401 based in liposomes^{50,72,96}, microspheres⁶², polymeric micelles⁶⁴ and nanoparticles^{51,97}. In most cases,
402 ATRA was encapsulated in the formulation^{50,72}, or chemically conjugated to the carrier⁵¹. This
403 effort was motivated by the fact that some of the patients treated with ATRA relapsed during the 4-
404 6 weeks treatment of daily oral administration. Follow-up studies indicated that resistance was due
405 to the decrease of ATRA concentration in the plasma, and thus inability of the drug to differentiate
406 leukemic cells⁴². The decrease was attributed to an induction of ATRA catabolism and increased
407 levels of RA-binding protein²⁰. Several ATRA delivery systems showed relative success in
408 reducing induction of ATRA catabolism and thus maintaining RA for longer periods in blood
409 plasma. For example, microspheres of poly(L-lactide) showed a nearly constant release rate of
410 ATRA for 5 weeks⁶². In addition, intravenous administration of RA-containing liposomes for 7
411 weeks in rats did not enhance RA catabolism⁵⁰. Maintenance of RA in the plasma reduced the
412 number of relapses⁹⁸.

413 Targeted delivery of RA-containing formulations to bone marrow has recently attracted
414 much attention⁹⁹. Bone marrow is the residence of hematopoietic stem cells that give rise to
415 myeloid and lymphoid cell lineages. In leukemia patients, hematopoietic stem cells are the target of
416 genetic mutations and thus aberrant activity. These altered cells are difficult to eliminate by
417 conventional antitumoral agents because drugs do not reach bone marrow at the required
418 concentration, and because the stem cell niche (a physical and functional entity that supports the
419 self-renewal and differentiation of stem cells) changes after chemotherapy, protecting altered
420 cells¹⁰⁰. RA is a potential treatment, combined with other drugs, for AML¹⁰¹ and chronic myeloid
421 leukemia¹⁰². In the last 10 years, a new therapeutic paradigm has emerged based in the targeted
422 delivery of formulations to bone marrow⁹⁹. Targeted delivery can be achieved by surface
423 modification of nanoparticles with bisphosphonate to promote binding to bone¹⁰³, by surface
424 modification of liposomes with folate to target the folate receptor, which is highly expressed in
425 AML cells^{21,104}, by the conjugation of nanoparticles with the surface of hematopoietic stem cells¹⁰⁵,
426 or by surface modification of nanoparticles with antibodies (e.g. CD45.1, CD117)^{106,107} or aptamers
427 (E-selectin thioaptamer)¹⁰⁸. Recently, we have developed a new platform to target bone marrow,
428 based on light-activatable nanoparticle formulations containing ATRA^{25,26}. The formulation was
429 highly internalized by leukemia-initiating cells and accumulated in the cell cytoplasm. Once
430 leukemia-initiating cells transfected with light-activatable nanoparticles containing RA were
431 administered intravenously in leukemic mice they tend to home in the bone marrow, in the
432 proximity of other leukemia cells. The irradiation of bone marrow with a blue light induced photo-
433 disassembly of the nanoparticles within the cells and consequently their differentiation. These cells

434 then secreted extracellular vesicles containing ATRA that interfered with cells supporting the stem
435 cell niche.

436

437 **7.1.3. Pre- and clinical applications**

438 Most RA-containing formulations are in clinical evaluation for treatment of blood tumors
439 (**Table 1**). Current challenges for translation of RA formulations to the clinic are related to two
440 factors: (i) capacity to incorporate multiple drugs besides RA; (ii) efficiently target hematopoietic
441 cells (**Figure 5**). In most cases, RA-based monotherapies are less efficient in interfering with
442 tumorigenesis and cancer progression than combination therapies²⁹. The efficiency of combination
443 therapies is likely ascribed to several factors: (i) a combination of drugs may act at different sites of
444 the anti-proliferative signaling pathway and thus be more effective in the inhibition process; (ii)
445 combination therapies may decrease the probability of cancer resistance; (iii) synergies between the
446 drugs reduces the dose necessary for therapy, as well as their toxicity and treatment time. Currently,
447 several clinical trials are investigating the combination of ATRA with arsenic trioxide, with arsenic
448 trioxide and gemtuzumab ozogamicin (a monoclonal antibody against CD33 conjugated with
449 ozogamicin, a cytotoxic agent), with epigenetic regulators such as tranylecypromine, with decitabine,
450 cytarabine and granulocyte-stimulating factor, among others, in the context of AML (**Table 1**).
451 Unfortunately, most RA formulations tested so far for blood cancers have not explored
452 simultaneous controlled release of RA and other agents. Thus, further investigation is needed to
453 address this issue. It should be noted that a liposomal formulation having 2 drugs (non-RA drugs)
454 has been approved recently for AML¹⁰⁹. Another important challenge in clinical translation of RA
455 formulations is targeting. Pre-clinical tests have addressed different leukemia cell targets, not yet
456 explored in clinical trials. The recent re-approval of gemtuzumab ozogamicin¹⁰⁹ may inspire new
457 approaches for RA formulation targeting.

458

459 **7.2. RA delivery systems for the treatment of solid tumors**

460 **7.2.1. RA mode of action**

461 One of the first applications of RA formulations for treatment of solid tumors was in
462 mild/moderate intraepithelial cervical neoplasia^{4,5}. ATRA was released by a collagen sponge with a
463 cervical cap at the tumor site by insertion in the cervix. A dose of ~0.4% of RA was selected for a
464 phase II trial. Fifty percent of the patients showed total regression of the disease^{4,5}. Systemic and
465 cervical side effects were mild and vaginal side effects were moderate and tolerable. Unfortunately,
466 the formulation did not reach the market, likely because it was not sufficient to reverse or suppress

467 more advanced dysplasia with acceptable local side effects in a phase III clinical trial¹¹⁰. The
468 antitumoral activity of RAs is linked to their differentiation and cell growth arrest properties¹¹¹. RA
469 has been tested in several clinical trials and the results showed that RA alone did not present
470 significant antitumoral activity against breast cancer¹¹²; however, when RA was combined with
471 tamoxifen or paclitaxel it showed moderate antitumoral activity^{113,114}.

472

473 **7.2.2. Type of RA-containing formulations**

474 In the last years, significant effort has been made by the scientific community to develop
475 formulations able to release RA and other neoplastic drugs. For example, nanoparticles containing
476 both ATRA and paclitaxel (to inhibit cell division because the cell cannot disrupt the polymerized
477 tubulin for cell division) had superior efficacy than formulations containing only paclitaxel⁵⁴. In
478 addition, pH-sensitive nanoparticles have been designed to release all-*trans* retinal and doxorubicin
479 in weakly acidic tumors (pH 6.5) or in acidic intracellular environments such as
480 endosomes/lysosomes (pH 4.5-5.5)¹¹⁵. All-*trans* retinal was chemically conjugated to the
481 nanoparticle polymer by hydrazone bonds that were labile under acidic conditions. Compared to
482 free drugs, the nanoparticle formulation increased the accumulation of the all-*trans*-retinal and
483 doxorubicin at the tumor site and induced higher levels of cell senescence and anti-tumoral activity.
484 Progress also has been made in targeting cancer-initiating cells, which are resistant to chemotherapy
485 and associated with tumor recurrence. ATRA liposomes and nanoparticles encapsulating
486 simultaneously ATRA and doxorubicin have been used successfully to arrest proliferation of breast
487 cancer-initiating cells and to differentiate them^{116,117}. In a combinatorial approach, RA
488 differentiated cancer-initiating cells, whereas antineoplastic agents, such as doxorubicin, killed non-
489 cancer initiating cells.

490

491 **7.2.3. Pre- and clinical applications**

492 The results of several clinical trials indicate that 13-*cis* RA, alone or in combination, seems
493 to be effective at different levels against malignant gliomas¹¹⁸, and cancer occurring in the central
494 and peripheral nervous system¹¹⁹. Moreover, ATRA inhibits proliferation of glioblastoma cells *in*
495 *vitro* and *in vivo*, while promoting their differentiation^{120,121}. With high concentrations, ATRA may
496 induce cell apoptosis¹²². RA-coated solid lipid nanoparticles showed higher *in vitro* toxicity
497 relatively to glioblastoma cells than the parent drug¹²³. In addition, polymeric micelles composed by
498 methoxy (mPEG)-grafted chitosan and encapsulating ATRA was more effective at inhibiting
499 glioblastoma cell line migration *in vitro* than the free agent⁶³.

500 Immune system plays an important role in modulating tumor progression; however, limited
501 infiltration of immune cells in the tumor site, as well as existence of immunosuppressive agents,
502 hamper their biological role. Therefore, a combination of chemotherapy with immunotherapy might
503 be a better strategy to fight tumor biology. Indeed, immunotherapy may increase sensitivity of
504 cancer cells to chemotherapy and thus reduce its side effects¹²⁴. Recently, a chemo-immunotherapy
505 approach for melanoma has been developed based on biodegradable hollow mesoporous
506 nanoparticles containing three drugs: doxorubicin, ATRA, and interleukin (IL)-2²³. IL-2 is a T cell
507 growth factor and thus can facilitate proliferation and activation of T cells. Animals treated with
508 formulations containing the three agents were the ones with the most effective tumor growth
509 inhibition and decreased metastasis. ATRA was effective in differentiating myeloid-derived
510 suppressor cells, and in synergy with doxorubicin, contributed to increase the number of dendritic
511 cells at the tumor site; IL-2 facilitated the proliferation of CD8⁺ T cells at the tumor site to promote
512 effective tumor killing.

513

514 **8. RA delivery systems for immune diseases**

515 **8.1. RA mode of action**

516 Inflammation is essential for the regulation of tissue homeostasis and barrier integrity (e.g.
517 BBB, mucosal barrier). However, when dysregulated, it contributes to the pathophysiology of many
518 diseases. In fact, retinoids have been described as potent anti-inflammatory and therefore protective
519 in pathologies such as chronic obstructive pulmonary disease⁷⁶, rheumatoid arthritis¹²⁵, psoriasis¹²⁶
520 and inflammatory bowel disease¹²⁷. At cellular levels, RA inhibited IL-6-driven induction of
521 proinflammatory Th₁₇ cells and promoted the differentiation of T_{reg} cells, which are important to
522 suppress excessive immune responses¹²⁸.

523 The role of RA in the gastrointestinal tract is particularly relevant since it is produced and
524 metabolized in the intestine, where it regulates the differentiation and function of diverse immune
525 cells, and supports mucosal barrier immunity¹²⁹. Indeed, ATRA regulates the activity of CD161⁺-
526 T_{reg} cells that support wound repair in intestinal mucosa¹⁴. These C-type lectin CD161 regulatory T
527 cells were found to induce cytokines that promoted epithelial barrier healing in the gut.
528 Accordingly, several studies have focused on the impact of vitamin A intake (i.e. RA obtained from
529 diet), both during development and in the adult. Accordingly, the levels of retinoids obtained from
530 the maternal diet increase the size of secondary lymphoid organs and the efficacy of adult immune
531 responses¹³, while the depletion of vitamin A, in the adult, led to a decrease of Th1, Th17, and ILC3
532 responses (a synonym of lowered immunity to bacterial infection), an effect which is counteracted
533 through time¹³⁰.

534

535 **8.2. Type of RA-containing formulations**

536 Several RA formulations have been developed to target immune cells and induce an
537 immunomodulatory response, such as solid lipid nanoparticles⁷⁶, polymeric nanoparticles⁵⁹,
538 nanostructured lipid carriers¹²⁷, among others. RA formulations with an average diameter of 130 to
539 250 nm^{59,76} and an entrapment efficiency of ATRA between 2.3 and 310 μg ¹²⁷ per mg of
540 nanoparticle have been developed. Macrophage phagocytosis was influenced largely by the
541 physicochemical properties of nanoparticles. For example, uptake is higher in nanoparticles with
542 high negative or positive surface charges¹³¹. In general, RA formulations showed sustained
543 intracellular RA delivery for a few days^{59,76}. RA formulations are taken up by macrophages and
544 induce anti-inflammatory responses by suppressing NF- κ B signaling and increasing bone
545 morphogenetic protein 2 signaling (pivotal for bone and cartilage development), as well as by
546 enhancing production of anti-inflammatory cytokines (e.g. IL-10)^{59,127}. In addition, RA
547 formulations have the capacity to enhance differentiation of naïve T cells to regulatory T cells¹²⁷.
548 The advantages of RA-containing nanoparticles *versus* free RA were demonstrated in inhibiting
549 expression of IL-6 and IL-8 in alveolar epithelial cells⁷⁶, but were not demonstrated in induction of
550 immune cell differentiation.

551

552 **8.3. Pre- and clinical applications**

553 Clinical trials using free RA in the context of immune thrombocytopenia (severe bleeding
554 disorder) and sclerosing cholangitis (inflammation and scarring of the bile ducts) are active and will
555 evaluate the role of RA as immunotherapy (**Table 1**). Current challenges for using RA formulations
556 for immune diseases are related to the *in vivo* demonstration of RA effects. This requires use of
557 formulations able to target specific cells, particularly immune cells and their surface receptors
558 (**Figure 5**). There are classes of molecules, including antibodies, carbohydrates, peptides, aptamers
559 that can be attached to RA formulations to target more specific immune cell populations.

560

561 **9. Future outlook**

562 The advantages of RA formulations compared to free RA have been demonstrated in various
563 biomedical applications. These are related to: (i) increased bioavailability; (ii) decreased toxicity
564 and skin irritation; (iii) increased *in vivo* half-life; (iv) increased RA photostability; (v) increased
565 cell targeting and capacity to cross biological barriers. Advantages of RA formulations have been
566 demonstrated in preclinical (in all applications described in this review) and clinical trials (cancer

567 and skin applications)^{73,132}. In the case of cancer, the high efficacy of RA formulations has been
568 attributed to a longer RA half-life. In fact, recent studies indicate that the antitumoral activity of RA
569 might be enhanced with formulations that extend the *in vivo* half-life of RA¹².

570 RA formulations have some limitations that preclude immediate translation into the clinic.
571 ATRA, 9-*cis*-RA and 13-*cis*-RA belong to the first generation of retinoids. They are being replaced
572 in some applications by third-generation retinoids¹³³. Although many formulations have been tested
573 in preclinical models, clinical translation of the formulations is relatively low. For example,
574 although nanoparticles delivered systemically have been tested in many animal tests, clinical
575 translation is relatively poor because of delivery (e.g. limited efficacy in terms of cell targeting,
576 limited capacity to cross biological barriers), technical (e.g. scale-up) and regulatory aspects (e.g.
577 study design and approval challenges). Use of antibodies, peptides, aptamers immobilized in the
578 surface of nanoparticles to target specific cellular receptors may accelerate clinical translation of
579 RA formulations. Indeed, the recent FDA approval of gemtuzumab ozogamicin (anti-CD33
580 antibody conjugated with a cytotoxic agent)¹⁰⁹ or daratumumab (anti-CD38 antibody)¹³⁴ might
581 accelerate clinical testing and approval of other formulations conjugated with antibodies. In
582 addition, nanoparticles/microparticles used in pre-clinical studies have been prepared in small
583 batches. Scale-up of these formulations is challenging, as they must comply with regulatory
584 guidelines, in terms of narrow size distribution, precise chemical composition and drug loading¹³⁵.
585 In the last years, progress has been made in producing more controlled formulations with size and
586 composition levels compatible with microfluidic systems¹³⁶. It is expected that these technological
587 advances might accelerate clinical translation of some RA formulations.

588 Despite progress in the last 50 years, many issues remain to be addressed. For example,
589 further preclinical and clinical studies are necessary to evaluate the mechanism of RA formulations
590 biodistribution, pharmacokinetics, clearance and toxicology. This requires development of
591 theranostic RA formulations that may be tracked *in vivo* by luminescence, magnetic resonance
592 imaging or positron emission tomography. Theranostic RA formulations are particularly relevant if
593 they are injected intravenously. Moreover, further formulations should be developed with capacity
594 to release multiple drugs in combination with RA. Pathologies discussed in this review (neurologic
595 diseases, cancer, immune diseases) are multifactorial and display complex signaling pathways and
596 symptoms. Although some progress has been made in the last 5 years regarding formulations with
597 capacity to release RA in combination with other agents^{55,115}, further effort is needed to better
598 control *in vivo* the half-life of each drug to match clinical dosage programs. Another area that
599 deserves further investigation is development of stimuli-responsive RA formulations. These
600 systems have ability to undergo physical and/or chemical changes in response to endogenous

601 biological or external triggers⁶⁵. The concept here is that the systems retain the drug and release it
602 after a specific trigger, enhancing its therapeutic efficacy and minimizing systemic toxicity. These
603 systems may be used for the targeted delivery of RA in hematopoietic²⁵ or neurogenic niches⁶¹.
604 Development of these systems requires development of linkers susceptible to pH or enzymes
605 present in the targeted cell/tissue. For example, several RA-polymer conjugates have been
606 developed to release RA by hydrolytic cleavage of ester bonds^{51,53}, by enzymatic cleavage of amide
607 bonds by proteases present in the targeted tissue^{28,65-67} or by reducing environments⁵⁷ such as the
608 cell cytoplasm. In addition, linkers may be cleaved by an external trigger, such as light with the
609 consequent release of RA^{25,26}.

610 RA is a very deeply conserved as a pathway and evolutionarily ancient. The overlap and
611 pleiotropic effects of RA is one of the key reasons for targeted delivery. It is expected that the
612 knowledge gathers in the development of retinoid formulations inspire others in the development of
613 formulations based in different drugs.

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Table 1- RA formulations tested in past and ongoing clinical trials.

Drug	Indication	Clinical trial
ATRA-containing liposomes	Acne	Phase I/II (NA) ⁷³
ATRA-containing collagen sponge	Mild/moderate intraepithelial cervical neoplasia	Phase II (NA) ^{4,5}
ATRA + pembrolizumab	Advanced melanoma	Phase I/Ib (NCT03200847)
ATRA + ipilimumab	Advanced melanoma	Phase II (NCT02403778)
13- <i>cis</i> RA + cabozantinib	Solid tumors	Phase I (NCT03611595)
13- <i>cis</i> RA + temozolomide + thiotepa + carboplatin	Brain tumor	Phase II (NCT00528437)
13- <i>cis</i> RA + 3F8/GM-CSF	Neuroblastoma	Phase II (NCT01183429)
13- <i>cis</i> RA + 3F8/GM-CSF	Neuroblastoma	Phase II (NCT01183897)
13- <i>cis</i> RA	Neuroblastoma	Phase I/II (NCT03291080)
13- <i>cis</i> RA + dinutuximab + lenalidomide	Neuroblastoma	Phase I (NCT01711554)
13- <i>cis</i> RA + several drugs	Neuroblastoma	NA (NCT01526603)
ATRA	Cholangitis, sclerosing	Phase II (NCT03359174)
ATRA + arsenic trioxide	APL	Phase II (NCT01404949)
ATRA + arsenic trioxide	APL	Phase III (NCT02339740)
ATRA + arsenic trioxide + gemtuzumab ozogamicin	APL	Phase II (NCT01409161)
ATRA + idarubicin	APL	NA (NCT01064557)
ATRA + arsenic trioxide + Realgar-Indigo naturalis formula	APL	Phase III (NCT02899169)
ATRA + several drugs	APL	Phase IV (NCT02200978)
ATRA + several drugs	APL	Phase III (NCT02688140)
ATRA + several drugs	APL	Phase III (NCT00482833)
ATRA	Acne vulgaris	Phase IV (NCT02620813)
ATRA	Multiple myeloma	Phase I/II (NCT02751255)
ATRA + rituximab	Immune thrombocytopenia	Phase II (NCT03304288)
ATRA + 5-azacitidine + lupron	Prostate cancer	Phase II (NCT03572387)
ATRA	Olfactory loss	NA (NCT03574701)
ATRA + tranlycypromine + cytarabine	AML	Phase I/II (NCT02717884)
ATRA + tranlycypromine	AML	Phase I (NCT02273102)
ATRA + gemtuzumab ozogamicin	AML	Phase III (NCT00893399)
ATRA + decitabine + cytarabine + G-CSF	AML	Phase II (NCT03356080)
ATRA + arsenic trioxide + cytarabine	AML	Phase I/II (NCT03031249)
ATRA + pioglitazone + azacitidine	AML	Phase II (NCT02942758)
ATRA + gemcitabine + Nab-paclitaxel	Pancreatic cancer	Phase I (NCT03307148)
ATRA + INCB059872+ azacitidine + nivolumab	Advanced malignancies	Phase I/II (NCT02712905)
13- <i>cis</i> RA + vorinostat + temozolamide	Glioblastoma	Phase I/II (NCT00555399)

Retinol + bakuchiol	Photoaging; wrinkles	Phase I/II (NCT03112863)
9- <i>cis</i> RA + cyclosporine A	Hand eczema	Phase III (NCT03026946)

644 APL= Acute promyelocytic leukemia; ATRA= all-*trans* retinoic acid; GM-CSF= Granulocyte-macrophage colony-
645 stimulating factor; NA= Not available.
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653 **Table 2- Synthetic retinoids in terminated or active (in bold) clinical trials and approved for commercialization.**

Drug	Receptor activity	Indication	Clinical trial
Tamibarotene (or Am80)	RAR α agonist	Crohn's disease	Phase II (NCT00417391)
		APL	Phase II (NCT00520208)
		Advanced non-small cell lung cancer	Phase I (NCT01337154)
Palovarotene	RAR γ agonist	AML or myelodysplastic syndrome	Phase II (NCT02807558)
		Eye dry disease	Phase I (CTP300302)
		Fibrodysplasia ossificans progressive	Phase III (NCT03312634)
Trifarotene (cream)	RAR γ agonist	Multiple osteochondromas	Phase II (NCT03442985)
		Moderate facial and truncal acne vulgaris	Phase III (NCT03915860)
		Autosomal recessive ichthyosis with lamellar scale	Phase II (NCT03738800)
Bexarotene (capsules)	Pan-agonist	Early cutaneous T-cell lymphoma	Phase I (NCT01804335)
		Refractory cutaneous T-cell lymphoma	Approved by FDA since 1999
		Tazarotene (gel or cream)	Pan-agonist
Adapalene (solution, cream and lotion)	Pan-agonist	Facial acne vulgaris	Approved by FDA since 1997
		Plaque psoriasis	Approved by FDA since 1997
		Acne	Approved by FDA since 1996
AGN194204	RXR agonist	Prostate cancer	Phase II (NCT01540071)
UAB30 / 9- <i>cis</i> -UAB30	RXR agonist	Non-melanoma skin cancer	Phase I/II (NCT03327064)
Fenretinide (oral powder and intravenous liquid emulsion)	Atypical retinoid	Peripheral T-cell lymphoma	Phase II (NCT02495415)
		Solid tumor (relapsed malignancies)	Phase I (NCT01553071)
		High risk cancer	Phase III (NCT01479192)
		Prevention of bladder cancer	Phase III (NCT00004154)
		Cervical neoplasia	Phase III (NCT00003075)
		Schizophrenia	Phase III (NCT00534898)
		Breast cancer	Phase III (NCT01357772)

654 APL= Acute promyelocytic leukemia; AML=Acute myeloid leukemia; FDA=Food Drug Administration.

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664 **Table 3-** Examples of formulations for the controlled release of RA in context of cancer, brain diseases, skin diseases,
665 immune diseases, stem cell differentiation, among others.
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Type	Diameter	Loading (µg/mg of carrier)	Delivery	Applications	Ref.
NPs	≈ 500 nm	≈ 76	Up to 3 h in the presence of trypsin	Pharmaceutical	97
NPs	< 200 nm	154	Release up to 10 days	Cancer	52
NPs	170-230 nm	20	80% cumulative drug release in 4 days	Immune diseases	59
NPs	170-185 nm	275	Sustained drug release observed over 72 h	Cancer	57
NPs	≈ 200 nm	86	17% cumulative RA release for 21 days	Stem cells and brain diseases	24,45,67
NPs	160 nm	150	50% release after 10 min irradiation	Cancer and brain diseases	25,61
NPs	≈ 100 nm	68-87	5 - 100% release of RA in 5 days	Stem cells and brain diseases	46
NPs	≈ 214 nm	30	38% cumulative release of RA in 48 h	Cancer	123
MPs	≈ 5.6 µm	80	Pseudo-zero order release for 5 weeks	Pharmaceutical	62
MPs	≈ 8 µm	3	Release up to 10 days	Stem cells	7
MPs	190 µm	57	Less than 3 h	Cancer	137
Mic	100-500 nm	40 - 130	NA	Pharmaceutical	49
Mic	100-400 nm	2.6	Parenteral administration; < 5% in 3 days	Cancer	64
Mic	50-200 nm	80% (w/w)	1-month delivery	Cancer	48
Lip	NA	NA	4.4 µg/mL blood <i>per</i> day (in humans)	Cancer	50,72,98
EFM	0.95-1.89 µm	5	80% cumulative RA release in 3.5 months	Stem cells	138
EFM	<5 µm	≈ 10	≈ 9.0 µM for 1 h	Stem cells	75
Scaffold	NA	≈ 20	Release over 8-28 h	Regenerative medicine	139
Scaffold	0.1-0.85 µm fibers	≈ 0.3 - 7	Sustained release of RA for up to 1 week	Stem cells	140

667 EFM= Electrospun fibrous mesh; h= hour; Lip= Liposome; Mic= Micelles; MPs= Microparticles; NA= Not available;

668 NPs= Nanoparticles; min= minute.

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686 **Table 4-** Marketed RA formulations.

Name/Company	Composition	Indication	Year approved	Ref.
9-cis RA (brand name: Panretin)/Eisai Inc.	Topical formulation; 0.05% or 0.1% gel containing alitretinoin	Cutaneous Kaposi's sarcoma; Treatment of recalcitrant chronic hand dermatitis	2000	141,142
13-cis RA (brand name: Accutane)/Roche	Topical formulation; 0.05% and 0.1% cream	Photoaging and acne	1982	142
ATRA	Topical formulation; 0.05% cream	Photoaging and acne	1971	2
Gel microsphere formulation containing ATRA/Advanced Polymer Systems	Topical formulation; macroporous beads, 10-25 µm in diameter	Acne	1997	77,143
9-cis RA	Topical formulation; 0.1% gel	AIDS-related Kaposi's sarcoma	1999	77
Retinol	Topical formulation; 0.5-5% lotion, cream	Cosmetic	NA	77
Retinaldehyde	Topical formulation; 0.01, 0.015, 0.1% cream	Cosmetic	NA	77
ATRA (brand name: Vesanoid)/Roche	Oral formulation of Tretinoin	Acute myeloid leukemia, in particular, acute promyelocytic leukemia	2000	134
Acitretin	Oral formulation	Psoriasis, disorders of keratinization	1997	144
13-cis RA	Oral formulation	Severe acne/related disorder	1982	77
Retinol	Oral formulation	Prevent/treat hypovitaminosis A	NA	77

687 ATRA= all-trans retinoic acid; NA= Not available

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References

- 1 Karrer, P., Morf, R. & Schopp, K. Information on vitamine A from train-oil. *Helv Chim Acta* **14**, 1035-1040 (1931).
- 2 Raza, K. *et al.* Nano-lipoidal carriers of tretinoin with enhanced percutaneous absorption, photostability, biocompatibility and anti-psoriatic activity. *Int J Pharm* **456**, 65-72, doi:10.1016/j.ijpharm.2013.08.019 (2013).
- 3 Breitman, T. R., Selonick, S. E. & Collins, S. J. Induction of differentiation of the human promyelocytic leukemia cell line (HL-60) by retinoic acid. *Proc Natl Acad Sci U S A* **77**, 2936-2940 (1980).
- 4 Meyskens, F. L., Jr. *et al.* A phase I trial of beta-all-trans-retinoic acid delivered via a collagen sponge and a cervical cap for mild or moderate intraepithelial cervical neoplasia. *J Natl Cancer Inst* **71**, 921-925 (1983).
- 5 Graham, V., Surwit, E. S., Weiner, S. & Meyskens, F. L., Jr. Phase II trial of beta-all-trans-retinoic acid for cervical intraepithelial neoplasia delivered via a collagen sponge and cervical cap. *West J Med* **145**, 192-195 (1986).
- 6 Chew, S. Y., Hufnagel, T. C., Lim, C. T. & Leong, K. W. Mechanical properties of single electrospun drug-encapsulated nanofibres. *Nanotechnology* **17**, 3880-3891, doi:10.1088/0957-4484/17/15/045 (2006).
- 7 Carpenedo, R. L. *et al.* Homogeneous and organized differentiation within embryoid bodies induced by microsphere-mediated delivery of small molecules. *Biomaterials* **30**, 2507-2515, doi:10.1016/j.biomaterials.2009.01.007 (2009).
- 8 Cabezas-Wallscheid, N. *et al.* Vitamin A-Retinoic Acid Signaling Regulates Hematopoietic Stem Cell Dormancy. *Cell* **169**, 807-823 e819, doi:10.1016/j.cell.2017.04.018 (2017).
- 9 Chanda, B., Ditadi, A., Iscove, N. N. & Keller, G. Retinoic acid signaling is essential for embryonic hematopoietic stem cell development. *Cell* **155**, 215-227, doi:10.1016/j.cell.2013.08.055 (2013).
- 10 Farinello, D. *et al.* A retinoic acid-dependent stroma-leukemia crosstalk promotes chronic lymphocytic leukemia progression. *Nat. Commun.* **9**, 1787, doi:10.1038/s41467-018-04150-7 (2018).
- 11 Kozono, S. *et al.* Arsenic targets Pin1 and cooperates with retinoic acid to inhibit cancer-driving pathways and tumor-initiating cells. *Nat. Commun.* **9**, 3069, doi:10.1038/s41467-018-05402-2 (2018).
- 12 Wei, S. *et al.* Active Pin1 is a key target of all-trans retinoic acid in acute promyelocytic leukemia and breast cancer. *Nature Medicine* **21**, 457-U230, doi:10.1038/nm.3839 (2015).
- 13 van de Pavert, S. A. *et al.* Maternal retinoids control type 3 innate lymphoid cells and set the offspring immunity. *Nature* **508**, 123-127, doi:10.1038/nature13158 (2014).
- 14 Povoleri, G. A. M. *et al.* Human retinoic acid-regulated CD161(+) regulatory T cells support wound repair in intestinal mucosa. *Nat Immunol* **19**, 1403-1414, doi:10.1038/s41590-018-0230-z (2018).
- 15 Johansson, H. J. *et al.* Retinoic acid receptor alpha is associated with tamoxifen resistance in breast cancer. *Nat. Commun.* **4**, 2175, doi:10.1038/ncomms3175 (2013).
- 16 Hashimoto-Hill, S. *et al.* RARalpha supports the development of Langerhans cells and langerin-expressing conventional dendritic cells. *Nat. Commun.* **9**, 3896, doi:10.1038/s41467-018-06341-8 (2018).
- 17 Chronopoulos, A. *et al.* ATRA mechanically reprograms pancreatic stellate cells to suppress matrix remodelling and inhibit cancer cell invasion. *Nat. Commun.* **7**, 12630, doi:10.1038/ncomms12630 (2016).
- 18 Szuts, E. Z. & Harosi, F. I. Solubility of retinoids in water. *Arch. Biochem. Biophys.* **287**, 297-304 (1991).
- 19 Ourique, A. F. *et al.* Improved photostability and reduced skin permeation of tretinoin: development of a semisolid nanomedicine. *Eur J Pharm Biopharm* **79**, 95-101, doi:10.1016/j.ejpb.2011.03.008 (2011).
- 20 Adamson, P. C. Pharmacokinetics of all-trans-retinoic acid: clinical implications in acute promyelocytic leukemia. *Semin Hematol* **31**, 14-17 (1994).
- 21 Pan, X. Q. *et al.* Strategy for the treatment of acute myelogenous leukemia based on folate receptor beta-targeted liposomal doxorubicin combined with receptor induction using all-trans retinoic acid. *Blood* **100**, 594-602 (2002).
- 22 Han, X. X. *et al.* Reversal of pancreatic desmoplasia by re-educating stellate cells with a tumour microenvironment-activated nanosystem. *Nature Communications* **9**, doi:ARTN 3390 10.1038/s41467-018-05906-x (2018).

766 23 Kong, M. *et al.* Biodegradable Hollow Mesoporous Silica Nanoparticles for Regulating Tumor
767 Microenvironment and Enhancing Antitumor Efficiency. *Theranostics* **7**, 3276-3292,
768 doi:10.7150/thno.19987 (2017).

769 24 Ferreira, R. *et al.* Retinoic acid-loaded polymeric nanoparticles enhance vascular regulation of neural
770 stem cell survival and differentiation after ischaemia. *Nanoscale* **8**, , 8126-8137, doi:10.1039/c5nr09077f
771 (2016).

772 25 Boto, C. *et al.* Prolonged intracellular accumulation of light-inducible nanoparticles in leukemia cells
773 allows their remote activation. *Nat Commun* **8**, 15204, doi:10.1038/ncomms15204 (2017).

774 26 Jimenez-Balsa, A. *et al.* Nanoparticles Conjugated with Photocleavable Linkers for the Intracellular
775 Delivery of Biomolecules. *Bioconjug Chem* **29**, 1485-1489, doi:10.1021/acs.bioconjchem.7b00820 (2018).

776 27 Morales, J. O., Valdes, K., Morales, J. & Oyarzun-Ampuero, F. Lipid nanoparticles for the topical
777 delivery of retinoids and derivatives. *Nanomedicine-Uk* **10**, 253-269, doi:10.2217/Nnm.14.159 (2015).

778 28 Duester, G. Retinoic acid synthesis and signaling during early organogenesis. *Cell* **134**, 921-931,
779 doi:10.1016/j.cell.2008.09.002 (2008).

780 29 Altucci, L., Leibowitz, M. D., Ogilvie, K. M., de Lera, A. R. & Gronemeyer, H. RAR and RXR
781 modulation in cancer and metabolic disease. *Nat Rev Drug Discov* **6**, 793-810, doi:10.1038/nrd2397 (2007).

782 30 Teng, X., Zhang, H., Snead, C. & Catravas, J. D. Molecular mechanisms of iNOS induction by IL-
783 1beta and IFN-gamma in rat aortic smooth muscle cells. *American Journal of Physiology - Cell Physiology*
784 **282**, C144-152 (2002).

785 31 di Masi, A. *et al.* Retinoic acid receptors: from molecular mechanisms to cancer therapy. *Mol*
786 *Aspects Med* **41**, 1-115, doi:10.1016/j.mam.2014.12.003 (2015).

787 32 Wagner, N., Benkali, K., Alio Saenz, A., Poncet, M. & Graeber, M. Clinical Pharmacology and
788 Safety of Trifarotene, a First-in-Class RARgamma-Selective Topical Retinoid. *J Clin Pharmacol*,
789 doi:10.1002/jcph.1566 (2020).

790 33 in *LiverTox: Clinical and Research Information on Drug-Induced Liver Injury* (2012).

791 34 Gaikwad, J., Sharma, S. & Hatware, K. V. Review on Characteristics and Analytical Methods of
792 Tazarotene: An Update. *Crit Rev Anal Chem* **50**, 90-96, doi:10.1080/10408347.2019.1586519 (2020).

793 35 Maden, M. Retinoic acid in the development, regeneration and maintenance of the nervous system.
794 *Nature Reviews Neuroscience* **8**, 755-765 (2007).

795 36 Schenk, T., Stengel, S. & Zelent, A. Unlocking the potential of retinoic acid in anticancer therapy. *Br*
796 *J Cancer* **111**, 2039-2045, doi:10.1038/bjc.2014.412 (2014).

797 37 Nunez, V. *et al.* Retinoid X receptor alpha controls innate inflammatory responses through the up-
798 regulation of chemokine expression. *Proc Natl Acad Sci U S A* **107**, 10626-10631,
799 doi:10.1073/pnas.0913545107 (2010).

800 38 Levin AA, B. T., Kazmer S, Grippo JF. 13-cis retinoic acid does not bind to retinoic acid receptors
801 alpha, beta and gamma. *Toxicologist* **12**, 181 (1992).

802 39 Al Tanoury, Z., Piskunov, A. & Rochette-Egly, C. Vitamin A and retinoid signaling: genomic and
803 nongenomic effects. *J Lipid Res* **54**, 1761-1775, doi:10.1194/jlr.R030833 (2013).

804 40 Fiorella, P. D. & Napoli, J. L. Microsomal retinoic acid metabolism. Effects of cellular retinoic acid-
805 binding protein (type I) and C18-hydroxylation as an initial step. *J Biol Chem* **269**, 10538-10544 (1994).

806 41 Stern, R. When a uniquely effective drug is teratogenic: the case of isotretinoin. *N Engl J Med* **320**,
807 1007-1009 (1989).

808 42 Muindi, J. *et al.* Continuous treatment with all-trans retinoic acid causes a progressive reduction in
809 plasma drug concentrations: implications for relapse and retinoid "resistance" in patients with acute
810 promyelocytic leukemia. *Blood* **79**, 299-303 (1992).

811 43 Zanotti, G., Dacunto, M. R., Malpeli, G., Folli, C. & Berni, R. Crystal-Structure of the Transthyretin
812 Retinoic-Acid Complex. *Eur J Biochem* **234**, 563-569, doi:DOI 10.1111/j.1432-1033.1995.563_b.x (1995).

813 44 Thunemann, A. F. & Beyermann, J. Polyethylenimine complexes with retinoic acid: Structure,
814 release profiles, and nanoparticles. *Macromolecules* **33**, 6878-6885, doi:DOI 10.1021/ma000416x (2000).

815 45 Maia, J. *et al.* Controlling the neuronal differentiation of stem cells by the intracellular delivery of
816 retinoic acid-loaded nanoparticles. *ACS nano* **5**, 97-106 (2010).

817 46 Zhang, R. *et al.* Traceable Nanoparticle Delivery of Small Interfering RNA and Retinoic Acid with
818 Temporally Release Ability to Control Neural Stem Cell Differentiation for Alzheimer's Disease Therapy.
819 *Adv Mater* **28**, 6345-6352, doi:10.1002/adma.201600554 (2016).

820 47 Giordano, G. G., Refojo, M. F. & Arroyo, M. H. Sustained delivery of retinoic acid from
821 microspheres of biodegradable polymer in PVR. *Invest Ophthalmol Vis Sci* **34**, 2743-2751 (1993).

822 48 Jeong, Y. I. *et al.* Polyion complex micelles composed of all-trans retinoic acid and poly (ethylene
823 glycol)-grafted-chitosan. *J Pharm Sci-US* **95**, 2348-2360, doi:10.1002/jps.20586 (2006).

824 49 Kim, D. G., Jeong, Y. I. & Nah, J. W. All-trans retinoic acid release from polyion-complex micelles
825 of methoxy poly(ethylene glycol) grafted chitosan. *J Appl Polym Sci* **105**, 3246-3254, doi:10.1002/app.26480
826 (2007).

827 50 Mehta, K., Sadeghi, T., Mcqueen, T. & Lopezberestein, G. Liposome Encapsulation Circumvents the
828 Hepatic-Clearance Mechanisms of All-Trans-Retinoic Acid. *Leukemia Research* **18**, 587-596, doi:Doi
829 10.1016/0145-2126(94)90040-X (1994).

830 51 Nam, Y. S. *et al.* Chemical immobilization of retinoic acid within poly(epsilon-caprolactone)
831 nanoparticles based on drug-polymer bioconjugates. *J Appl Polym Sci* **89**, 1631-1637,
832 doi:10.1002/app.12366 (2003).

833 52 Kim, D. G. *et al.* All-trans retinoic acid-associated low molecular weight water-soluble chitosan
834 nanoparticles based on ion complex. *Macromol Res* **14**, 66-72, doi:Doi 10.1007/Bf03219070 (2006).

835 53 Castleberry, S. A., Quadir, M. A., Abu Sharkh, M., Shopsowitz, K. E. & Hammond, P. T. Polymer
836 conjugated retinoids for controlled transdermal delivery. *Journal of Controlled Release* **262**, 1-9,
837 doi:10.1016/j.jconrel.2017.07.003 (2017).

838 54 Hou, L., Yao, J., Zhou, J. P. & Zhang, Q. Pharmacokinetics of a paclitaxel-loaded low molecular
839 weight heparin-all-trans-retinoid acid conjugate ternary nanoparticulate drug delivery system. *Biomaterials*
840 **33**, 5431-5440, doi:10.1016/j.biomaterials.2012.03.070 (2012).

841 55 Yao, J., Zhang, L., Zhou, J. P., Liu, H. P. & Zhang, Q. Efficient Simultaneous Tumor Targeting
842 Delivery of All-Trans Retinoid Acid and Paclitaxel Based on Hyaluronic Acid-Based Multifunctional
843 Nanocarrier. *Mol Pharmaceut* **10**, 1080-1091, doi:10.1021/mp3005808 (2013).

844 56 Park, K. M. *et al.* All-trans-retinoic acid (ATRA)-grafted polymeric gene carriers for nuclear
845 translocation and cell growth control. *Biomaterials* **30**, 2642-2652, doi:10.1016/j.biomaterials.2009.01.025
846 (2009).

847 57 Huang, H. *et al.* Co-delivery of all-trans-retinoic acid enhances the anti-metastasis effect of albumin-
848 bound paclitaxel nanoparticles. *Chem Commun* **53**, 212-215, doi:10.1039/c6cc08146k (2017).

849 58 Cho, C. S. *et al.* Receptor-mediated delivery of all trans-retinoic acid to hepatocyte using poly(L-
850 lactic acid) nanoparticles coated with galactose-carrying polystyrene. *J Control Release* **77**, 7-15, doi:Doi
851 10.1016/S0168-3659(01)00390-X (2001).

852 59 Almouazen, E. *et al.* Development of a nanoparticle-based system for the delivery of retinoic acid
853 into macrophages. *Int J Pharmaceut* **430**, 207-215, doi:10.1016/j.ijpharm.2012.03.025 (2012).

854 60 Park, S. J. *et al.* Highly Efficient and Rapid Neural Differentiation of Mouse Embryonic Stem Cells
855 Based on Retinoic Acid Encapsulated Porous Nanoparticle. *Acs Appl Mater Inter* **9**, 34634-34640,
856 doi:10.1021/acsami.71309760 (2017).

857 61 Santos, T. *et al.* Blue light potentiates neurogenesis induced by retinoic acid-loaded responsive
858 nanoparticles. *Acta Biomater* **59**, 293-302, doi:10.1016/j.actbio.2017.06.044 (2017).

859 62 Choi, Y. *et al.* Long-term delivery of all-trans-retinoic acid using biodegradable PLLA/PEG-PLLA
860 blended microspheres. *Int J Pharm* **215**, 67-81, doi:Doi 10.1016/S0378-5173(00)00676-1 (2001).

861 63 Jeong, Y. I. *et al.* Polyion complex micelles composed of all-trans retinoic acid and poly (ethylene
862 glycol)-grafted-chitosan. *J Pharm Sci* **95**, 2348-2360, doi:10.1002/jps.20586 (2006).

863 64 Zuccari, G., Carosio, R., Fini, A., Montaldo, P. G. & Orienti, I. Modified polyvinylalcohol for
864 encapsulation of all-trans-retinoic acid in polymeric micelles. *J Control Release* **103**, 369-380,
865 doi:10.1016/j.jconrel.20004.12.016 (2005).

866 65 Mura, S., Nicolas, J. & Couvreur, P. Stimuli-responsive nanocarriers for drug delivery. *Nat Mater*
867 **12**, 991-1003, doi:10.1038/nmat3776 (2013).

868 66 Schug, T. T., Berry, D. C., Shaw, N. S., Travis, S. N. & Noy, N. Opposing effects of retinoic acid on
869 cell growth result from alternate activation of two different nuclear receptors. *Cell* **129**, 723-733,
870 doi:10.1016/j.cell.2007.02.050 (2007).

871 67 Santos, T. *et al.* Polymeric nanoparticles to control the differentiation of neural stem cells in the
872 subventricular zone of the brain. *ACS Nano* **6**, 10463-10474, doi:10.1021/nn304541h (2012).

873 68 Muindi, J. R. *et al.* Clinical pharmacology of oral all-trans retinoic acid in patients with acute
874 promyelocytic leukemia. *Cancer Res* **52**, 2138-2142 (1992).

875 69 Esteves, M. *et al.* Retinoic acid-loaded polymeric nanoparticles induce neuroprotection in a mouse
876 model for Parkinson's disease. *Front. Aging Neurosci.* **7**, 20, doi:10.3389/fnagi.2015.00020 (2015).

877 70 Nastruzzi, C., Walde, P., Menegatti, E. & Gambari, R. Liposome-associated retinoic acid. Increased
878 in vitro antiproliferative effects on neoplastic cells. *FEBS Lett* **259**, 293-296, doi:10.1016/0014-
879 5793(90)80030-m (1990).

880 71 Maia, J. *et al.* Controlling the neuronal differentiation of stem cells by the intracellular delivery of
881 retinoic acid-loaded nanoparticles. *ACS Nano* **5**, 97-106, doi:10.1021/nn101724r (2011).

882 72 Estey, E. *et al.* Alterations in tretinoin pharmacokinetics following administration of liposomal all-
883 trans retinoic acid. *Blood* **87**, 3650-3654 (1996).

884 73 Rahman, S. A. *et al.* Tretinoin-loaded liposomal formulations: from lab to comparative clinical study
885 in acne patients. *Drug Deliv* **23**, 1184-1193, doi:10.3109/10717544.2015.1041578 (2016).

886 74 Janesick, A., Wu, S. C. & Blumberg, B. Retinoic acid signaling and neuronal differentiation. *Cell*
887 *Mol Life Sci* **72**, 1559-1576, doi:10.1007/s00018-014-1815-9 (2015).

888 75 Tzezana, R., Reznik, S., Blumenthal, J., Zussman, E. & Levenberg, S. Regulation of stem cell
889 differentiation by control of retinoic acid gradients in hydrospun 3D scaffold. *Macromol Biosci* **12**, 598-607,
890 doi:10.1002/mabi.201100312 (2012).

891 76 Payne, C. M. *et al.* Evaluation of the Immunomodulatory Effects of All-Trans Retinoic Acid Solid
892 Lipid Nanoparticles and Human Mesenchymal Stem Cells in an A549 Epithelial Cell Line Model. *Pharm*
893 *Res* **36**, 50, doi:10.1007/s11095-019-2583-x (2019).

894 77 Sardana, K. & Sehgal, V. N. Retinoids: fascinating up-and-coming scenario. *J Dermatol* **30**, 355-380
895 (2003).

896 78 Saiag, P. *et al.* Treatment of early AIDS-related Kaposi's sarcoma with oral all-trans-retinoic acid:
897 results of a sequential non-randomized phase II trial. *Aids* **12**, 2169-2176, doi:10.1097/00002030-
898 199816000-00012 (1998).

899 79 Castro, G. A. *et al.* Formation of ion pairing as an alternative to improve encapsulation and stability
900 and to reduce skin irritation of retinoic acid loaded in solid lipid nanoparticles. *Int J Pharm* **381**, 77-83,
901 doi:10.1016/j.ijpharm.2009.07.025 (2009).

902 80 Masini, V., Bonte, F., Meybeck, A. & Wepierre, J. Cutaneous bioavailability in hairless rats of
903 tretinoin in liposomes or gel. *J Pharm Sci* **82**, 17-21 (1993).

904 81 Mehnert, W. & Mader, K. Solid lipid nanoparticles Production, characterization and applications.
905 *Adv Drug Deliver Rev* **64**, 83-101, doi:10.1016/j.addr.2012.09.021 (2012).

906 82 Ourique, A. F., Pohlmann, A. R., Guterres, S. S. & Beck, R. C. R. Tretinoin-loaded nanocapsules:
907 Preparation, physicochemical characterization, and photostability study. *Int J Pharmaceut* **352**, 1-4,
908 doi:10.1016/j.ijpharm.2007.12.035 (2008).

909 83 Laptewa, M., Moller, M., Gurny, R. & Kalia, Y. N. Self-assembled polymeric nanocarriers for the
910 targeted delivery of retinoic acid to the hair follicle. *Nanoscale* **7**, 18651-18662, doi:10.1039/c5nr04770f
911 (2015).

912 84 Xue, J. *et al.* Neutrophil-mediated anticancer drug delivery for suppression of postoperative
913 malignant glioma recurrence. *Nat Nanotechnol* **12**, 692-700, doi:10.1038/nnano.2017.54 (2017).

914 85 Zlokovic, B. V. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other
915 disorders. *Nat. Rev. Neurosci.* **12**, 723-738, doi:10.1038/nrn3114 (2011).

916 86 Ding, Y. *et al.* Retinoic acid attenuates beta-amyloid deposition and rescues memory deficits in an
917 Alzheimer's disease transgenic mouse model. *J Neurosci* **28**, 11622-11634, doi:10.1523/JNEUROSCI.3153-
918 08.2008 (2008).

919 87 Chakrabarti, M. *et al.* Molecular Signaling Mechanisms of Natural and Synthetic Retinoids for
920 Inhibition of Pathogenesis in Alzheimer's Disease. *J Alzheimers Dis* **50**, 335-352, doi:10.3233/JAD-150450
921 (2016).

922 88 Zhang, R. *et al.* Traceable nanoparticle delivery of small interfering RNA and retinoic acid with
923 temporally release ability to control neural stem cell differentiation for alzheimer's disease therapy.
924 *Advanced Materials* **28**, 6345-6352 (2016).

925 89 McCaffery, P. & Dräger, U. High levels of a retinoic acid-generating dehydrogenase in the meso-
926 telencephalic dopamine system. *Proceedings of the National Academy of Sciences* **91**, 7772-7776 (1994).

927 90 Jankovic, J., Chen, S. & Le, W. The role of Nurr1 in the development of dopaminergic neurons and
928 Parkinson's disease. *Progress in neurobiology* **77**, 128-138 (2005).

929 91 Smits, S. M., Ponnio, T., Conneely, O. M., Burbach, J. P. H. & Smidt, M. P. Involvement of Nurr1 in
930 specifying the neurotransmitter identity of ventral midbrain dopaminergic neurons. *European Journal of*
931 *Neuroscience* **18**, 1731-1738 (2003).

932 92 Islam, M. M. & Mohamed, Z. Computational and Pharmacological Target of Neurovascular Unit for
933 Drug Design and Delivery. *Biomed Res Int* **2015**, 731292, doi:10.1155/2015/731292 (2015).

934 93 Mizze, M. R. *et al.* Retinoic acid induces blood-brain barrier development. *J. Neurosci.* **33**, 1660-
935 1671, doi:10.1523/JNEUROSCI.1338-12.2013 (2013).

936 94 Machado-Pereira, M., Santos, T., Ferreira, L., Bernardino, L. & Ferreira, R. Intravenous
937 administration of retinoic acid-loaded polymeric nanoparticles prevents ischemic injury in the immature
938 brain. *Neurosci. Lett.* **673**, 116-121, doi:10.1016/j.neulet.2018.02.066 (2018).

939 95 Burnett, A. K. *et al.* Arsenic trioxide and all-trans retinoic acid treatment for acute promyelocytic
940 leukaemia in all risk groups (AML17): results of a randomised, controlled, phase 3 trial. *Lancet Oncol* **16**,
941 1295-1305, doi:10.1016/S1470-2045(15)00193-X (2015).

942 96 Drach, J., Lopezberestein, G., Mcqueen, T., Andreeff, M. & Mehta, K. Induction of Differentiation
943 in Myeloid-Leukemia Cell-Lines and Acute Promyelocytic Leukemia-Cells by Liposomal All-Trans-
944 Retinoic Acid. *Cancer Res* **53**, 2100-2104 (1993).

945 97 Ezpeleta, I. *et al.* Gliadin nanoparticles for the controlled release of all-trans-retinoic acid. *Int J*
946 *Pharmaceut* **131**, 191-200, doi:Doi 10.1016/0378-5173(95)04338-1 (1996).

947 98 Estey, E. H. *et al.* Molecular remissions induced by liposomal-encapsulated all-trans retinoic acid in
948 newly diagnosed acute promyelocytic leukemia. *Blood* **94**, 2230-2235 (1999).

949 99 Mu, C. F. *et al.* Targeted drug delivery for tumor therapy inside the bone marrow. *Biomaterials* **155**,
950 191-202, doi:10.1016/j.biomaterials.2017.11.029 (2018).

951 100 Duan, C. W. *et al.* Leukemia Propagating Cells Rebuild an Evolving Niche in Response to Therapy.
952 *Cancer Cell* **25**, 778-793, doi:10.1016/j.ccr.2014.04.015 (2014).

953 101 Montesinos, P. *et al.* Differentiation syndrome in patients with acute promyelocytic leukemia treated
954 with all-trans retinoic acid and anthracycline chemotherapy: characteristics, outcome, and prognostic factors.
955 *Blood* **113**, 775-783, doi:10.1182/blood-2008-07-168617 (2009).

956 102 Russo, D. *et al.* All-trans retinoic acid (ATRA) in patients with chronic myeloid leukemia in the
957 chronic phase. *Leukemia* **12**, 449-454, doi:DOI 10.1038/sj.leu.2400988 (1998).

958 103 Swami, A. *et al.* Engineered nanomedicine for myeloma and bone microenvironment targeting. *Proc*
959 *Natl Acad Sci U S A* **111**, 10287-10292, doi:10.1073/pnas.1401337111 (2014).

960 104 Lu, Y. *et al.* Role of formulation composition in folate receptor-targeted liposomal doxorubicin
961 delivery to acute myelogenous leukemia cells. *Mol Pharm* **4**, 707-712, doi:10.1021/mp0700581 (2007).

962 105 Stephan, M. T., Moon, J. J., Um, S. H., Bershteyn, A. & Irvine, D. J. Therapeutic cell engineering
963 with surface-conjugated synthetic nanoparticles. *Nat Med* **16**, 1035-1041, doi:10.1038/nm.2198 (2010).

964 106 Dorrance, A. M. *et al.* Targeting leukemia stem cells in vivo with antagonomiR-126 nanoparticles in
965 acute myeloid leukemia. *Leukemia* **29**, 2143-2153, doi:10.1038/leu.2015.139 (2015).

966 107 Barth, B. M. *et al.* Targeted indocyanine-green-loaded calcium phosphosilicate nanoparticles for in
967 vivo photodynamic therapy of leukemia. *ACS Nano* **5**, 5325-5337, doi:10.1021/nn2005766 (2011).

968 108 Zong, H. *et al.* In vivo targeting of leukemia stem cells by directing parthenolide-loaded
969 nanoparticles to the bone marrow niche. *Leukemia* **30**, 1582-1586, doi:10.1038/leu.2015.343 (2016).

970 109 Abdel-Wahab, O. A Landmark Year for FDA-Approved Therapies for Acute Myeloid Leukemia.
971 *Blood* **15** (2018).

972 110 Meyskens, F. L. *et al.* Enhancement of Regression of Cervical Intraepithelial Neoplasia-Ii (Moderate
973 Dysplasia) with Topically Applied All-Trans-Retinoic Acid - a Randomized Trial. *J Natl Cancer I* **86**, 539-
974 543, doi:DOI 10.1093/jnci/86.7.539 (1994).

975 111 Hua, S. J., Kittler, R. & White, K. P. Genomic Antagonism between Retinoic Acid and Estrogen
976 Signaling in Breast Cancer. *Cell* **137**, 1259-1271, doi:10.1016/j.cell.2009.04.043 (2009).

977 112 Sutton, L. M., Warmuth, M. A., Petros, W. P. & Winer, E. P. Pharmacokinetics and clinical impact
978 of all-trans retinoic acid in metastatic breast cancer: a phase II trial. *Cancer Chemother Pharmacol* **40**, 335-
979 341, doi:10.1007/s002800050666 (1997).

980 113 Budd, G. T. *et al.* Phase I/II trial of all-trans retinoic acid and tamoxifen in patients with advanced
981 breast cancer. *Clin Cancer Res* **4**, 635-642 (1998).

982 114 Bryan, M. *et al.* A pilot phase II trial of all-trans retinoic acid (Vesanoid) and paclitaxel (Taxol) in
983 patients with recurrent or metastatic breast cancer. *Invest New Drugs* **29**, 1482-1487, doi:10.1007/s10637-
984 010-9478-3 (2011).

985 115 Zhang, Y. *et al.* Retinal-conjugated pH-sensitive micelles induce tumor senescence for boosting
986 breast cancer chemotherapy. *Biomaterials* **83**, 219-232, doi:10.1016/j.biomaterials.2016.01.023 (2016).

987 116 Li, R. J. *et al.* All-trans retinoic acid stealth liposomes prevent the relapse of breast cancer arising
988 from the cancer stem cells. *J Control Release* **149**, 281-291, doi:10.1016/j.jconrel.2010.10.019 (2011).

989 117 Sun, R. *et al.* Co-delivery of all-trans-retinoic acid and doxorubicin for cancer therapy with
990 synergistic inhibition of cancer stem cells. *Biomaterials* **37**, 405-414, doi:10.1016/j.biomaterials.2014.10.018
991 (2015).

992 118 See, S. J., Levin, V. A., Yung, W. K., Hess, K. R. & Groves, M. D. 13-cis-retinoic acid in the
993 treatment of recurrent glioblastoma multiforme. *Neuro Oncol* **6**, 253-258, doi:10.1215/S1152851703000607
994 (2004).

995 119 Matthay, K. K. *et al.* Long-term results for children with high-risk neuroblastoma treated on a
996 randomized trial of myeloablative therapy followed by 13-cis-retinoic acid: a children's oncology group
997 study. *J. Clin. Oncol.* **27**, 1007-1013, doi:10.1200/JCO.2007.13.8925 (2009).

998 120 Ying, M. *et al.* Regulation of glioblastoma stem cells by retinoic acid: role for Notch pathway
999 inhibition. *Oncogene* **30**, 3454-3467, doi:10.1038/onc.2011.58 (2011).

1000 121 Niu, C. S. *et al.* Effect of all-trans retinoic acid on the proliferation and differentiation of brain tumor
1001 stem cells. *J. Exp. Clin. Cancer Res.* **29**, 113, doi:10.1186/1756-9966-29-113 (2010).

1002 122 Karsy, M., Albert, L., Tobias, M. E., Murali, R. & Jhanwar-Uniyal, M. All-trans retinoic acid
1003 modulates cancer stem cells of glioblastoma multiforme in an MAPK-dependent manner. *Anticancer Res.*
1004 **30**, 4915-4920 (2010).

1005 123 Liu, J. L. *et al.* Preparation of N, N, N-trimethyl chitosan-functionalized retinoic acid-loaded lipid
1006 nanoparticles for enhanced drug delivery to glioblastoma. *Trop J Pharm Res* **16**, 1765-1772,
1007 doi:10.4314/tjpr.v16i8.3 (2017).

1008 124 Wang, W. *et al.* Effector T Cells Abrogate Stroma-Mediated Chemoresistance in Ovarian Cancer.
1009 *Cell* **165**, 1092-1105, doi:10.1016/j.cell.2016.04.009 (2016).

1010 125 Nozaki, Y. *et al.* Anti-inflammatory effect of all-trans-retinoic acid in inflammatory arthritis. *Clin*
1011 *Immunol* **119**, 272-279, doi:10.1016/j.clim.2005.11.012 (2006).

1012 126 Fredriksson, T. & Pettersson, U. Severe Psoriasis - Oral Therapy with a New Retinoid.
1013 *Dermatologica* **157**, 238-244, doi:Doi 10.1159/000250839 (1978).

1014 127 Zai, K. *et al.* Regulation of inflammatory response of macrophages and induction of regulatory T
1015 cells by using retinoic acid-loaded nanostructured lipid carrier. *J Biomater Sci Polym Ed* **30**, 1-11,
1016 doi:10.1080/09205063.2018.1493671 (2019).

1017 128 Mucida, D. *et al.* Reciprocal T(H)17 and regulatory T cell differentiation mediated by retinoic acid.
1018 *Science* **317**, 256-260, doi:10.1126/science.1145697 (2007).

1019 129 Erkelens, M. N. & Mebius, R. E. Retinoic Acid and Immune Homeostasis: A Balancing Act. *Trends*
1020 *Immunol* **38**, 168-180, doi:10.1016/j.it.2016.12.006 (2017).

1021 130 Spencer, S. P. *et al.* Adaptation of innate lymphoid cells to a micronutrient deficiency promotes type
1022 2 barrier immunity. *Science* **343**, 432-437, doi:10.1126/science.1247606 (2014).

1023 131 He, C. B., Hu, Y. P., Yin, L. C., Tang, C. & Yin, C. H. Effects of particle size and surface charge on
1024 cellular uptake and biodistribution of polymeric nanoparticles. *Biomaterials* **31**, 3657-3666,
1025 doi:10.1016/j.biomaterials.2010.01.065 (2010).

1026 132 Jain, P. *et al.* Single-agent liposomal all-trans-retinoic Acid as initial therapy for acute promyelocytic
1027 leukemia: 13-year follow-up data. *Clin Lymphoma Myeloma Leuk* **14**, e47-49,
1028 doi:10.1016/j.clml.2013.08.004 (2014).

1029 133 Leyden, J., Stein-Gold, L. & Weiss, J. Why Topical Retinoids Are Mainstay of Therapy for Acne.
1030 *Dermatology Ther* **7**, 293-304, doi:10.1007/s13555-017-0185-2 (2017).

1031 134 Deshantri, A. K. *et al.* Nanomedicines for the treatment of hematological malignancies. *J Control*
1032 *Release* **287**, 194-215, doi:10.1016/j.jconrel.2018.08.034 (2018).

1033 135 Anselmo, A. C. & Mitragotri, S. Nanoparticles in the clinic. *Bioeng Transl Med* **1**, 10-29,
1034 doi:10.1002/btm2.10003 (2016).

1035 136 Karnik, R. *et al.* Microfluidic platform for controlled synthesis of polymeric nanoparticles. *Nano Lett*
1036 **8**, 2906-2912, doi:10.1021/nl801736q (2008).

- 1037 137 Shelley, R. S., Jun, H. W., Price, J. C. & Cadwallader, D. E. Blood Level Studies of All-Trans-
1038 Retinoic and 13-Cis-Retinoic Acids in Rats Using Different Formulations. *J Pharm Sci-US* **71**, 904-907,
1039 doi:DOI 10.1002/jps.2600710816 (1982).
- 1040 138 Puppi, D., Piras, A. M., Detta, N., Dinucci, D. & Chiellini, F. Poly(lactic-co-glycolic acid)
1041 electrospun fibrous meshes for the controlled release of retinoic acid. *Acta Biomater* **6**, 1258-1268,
1042 doi:10.1016/j.actbio.2009.08.015 (2010).
- 1043 139 O'Leary, C., O'Brien, F. J. & Cryan, S. A. Retinoic Acid-Loaded Collagen-Hyaluronate Scaffolds: A
1044 Bioactive Material for Respiratory Tissue Regeneration. *Acs Biomater Sci Eng* **3**, 1381-1393,
1045 doi:10.1021/acsbomaterials.6b00561 (2017).
- 1046 140 Damanik, F. F. R., van Blitterswijk, C., Rotmans, J. & Moroni, L. Enhancement of synthesis of
1047 extracellular matrix proteins on retinoic acid loaded electrospun scaffolds. *J Mater Chem B* **6**, 6468-6480,
1048 doi:10.1039/c8tb01244j (2018).
- 1049 141 Mukherjee, S. *et al.* Retinoids in the treatment of skin aging: an overview of clinical efficacy and
1050 safety. *Clin Interv Aging* **1**, 327-348 (2006).
- 1051 142 Riahi, R. R., Bush, A. E. & Cohen, P. R. Topical Retinoids: Therapeutic Mechanisms in the
1052 Treatment of Photodamaged Skin. *Am J Clin Dermatol* **17**, 265-276, doi:10.1007/s40257-016-0185-5 (2016).
- 1053 143 Berger, R. *et al.* Tretinoin gel microspheres 0.04% versus 0.1% in adolescents and adults with mild
1054 to moderate acne vulgaris: A 12-week, multicenter, randomized, double-blind, parallel-group, phase IV trial.
1055 *Clin Ther* **29**, 1086-1097, doi:10.1016/j.clinthera.2007.06.021 (2007).
- 1056 144 Kligman, A. M. The growing importance of topical retinoids in clinical dermatology: a retrospective
1057 and prospective analysis. *J Am Acad Dermatol* **39**, S2-S7, doi:Doi 10.1016/S0190-9622(98)70437-2 (1998).

1058

1059

1060 Captions

1061

1062 **Figure 1- Milestones in RA formulations research.**

1063

1064 **Figure 2- Retinoid chemical structures (a) and RA signaling pathway (b).** Blood-circulating
1065 retinol is internalized through membrane transporter and receptor *stimulated by retinoic acid 6*
1066 (STRA6) and converted into all-*trans* RA, which binds to cellular retinoic-acid-binding protein type
1067 2 for signaling in the nucleus. RA triggers gene transcription by binding to RA receptors (RAR) and
1068 to the retinoid X receptor (RXR). In the presence of the ligand, RAR and RXR heterodimerize on
1069 retinoic acid-response element (RARE) sequences located in promoter regions inducing the
1070 transcription of target genes. Of note, RXR may also homodimerize and trigger gene transcription
1071 (not depicted in the illustration). RA signaling may also occur via activation of receptors associated
1072 with lipid rafts located on the cell surface, which trigger transcriptional activation of target genes by
1073 histone and receptor phosphorylation in the cell nucleus. CRBP= cellular retinol-binding protein 1;
1074 MAPK= mitogen activated protein kinases; P = phosphorylation; RAL = retinaldehyde; RALDH=
1075 retinaldehyde dehydrogenase; RBP4 = retinol-binding protein 4; RDH = retinol dehydrogenase.

1076

1077 **Figure 3- Main outcomes of RA-based therapies in pathological contexts.** AD= Alzheimer's
1078 disease; PD= Parkinson's disease.

1079

1080 **Figure 4- Challenges and advances in topical and systemic administration of RA-containing**
1081 **formulations.** Challenges include: (a) crossing endothelial barriers for extravasation of RA-
1082 containing formulations into a specific body region; (b) targeting of RA-containing formulations to
1083 specific cells; (c) development of formulations that combine RA with other pharmacological agents,
1084 able to release each agent with a specific release kinetics. Advances include (a) development of
1085 formulations with less toxicity; (b) release of RA with variable release kinetics to achieve variable
1086 biological action; (c) action mechanism of RA during development and disease; (d) use of cells to
1087 transport RA-containing formulations to specific regions in the body followed by the triggering of
1088 the formulations by intrinsic (e.g. temperature, pH) or extrinsic (e.g. ultrasound, light) stimuli.

1089

1090

Figure 1

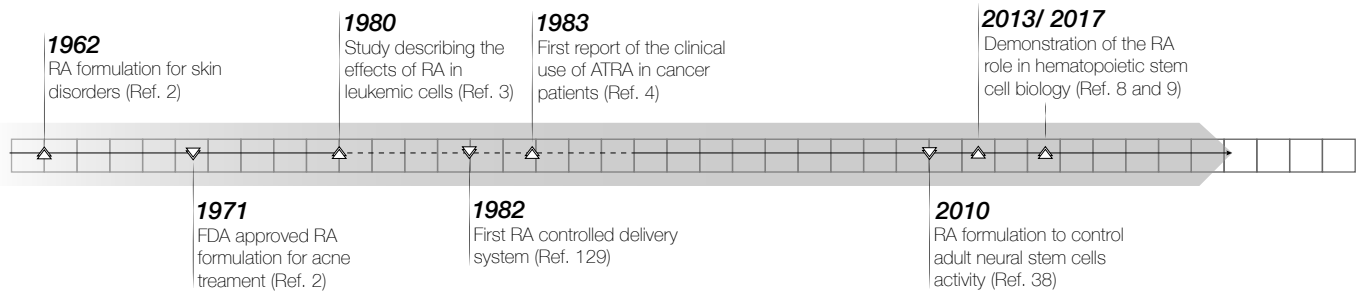


Figure 2

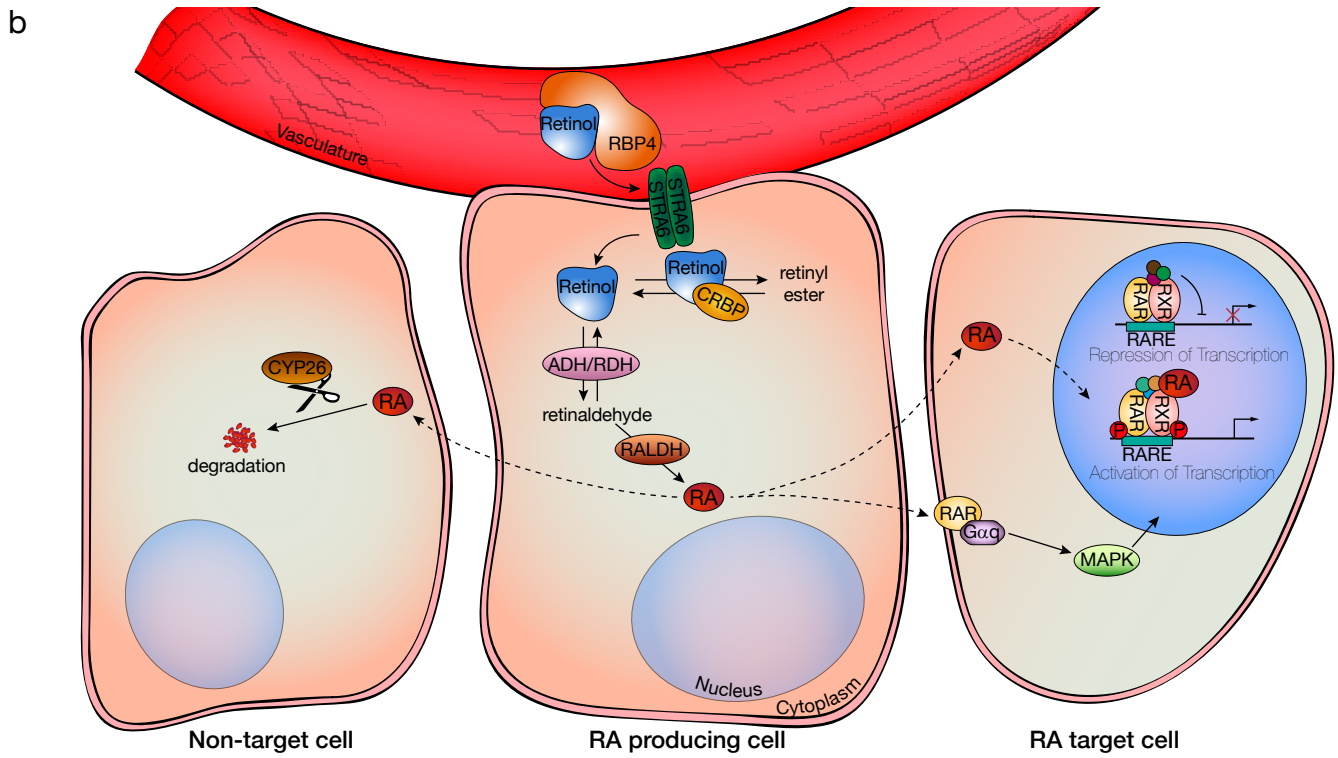
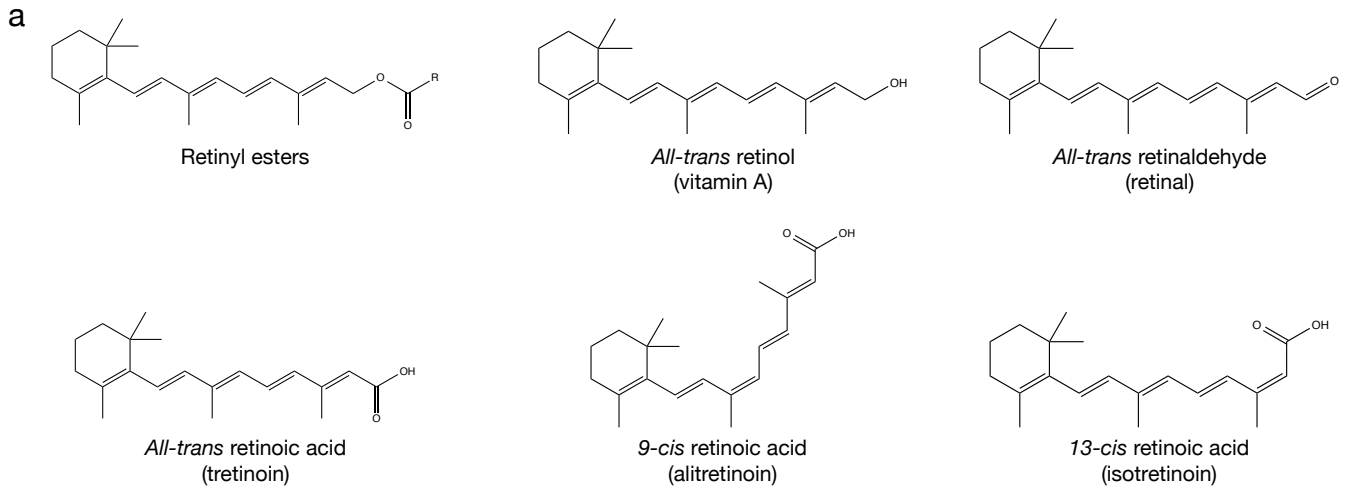
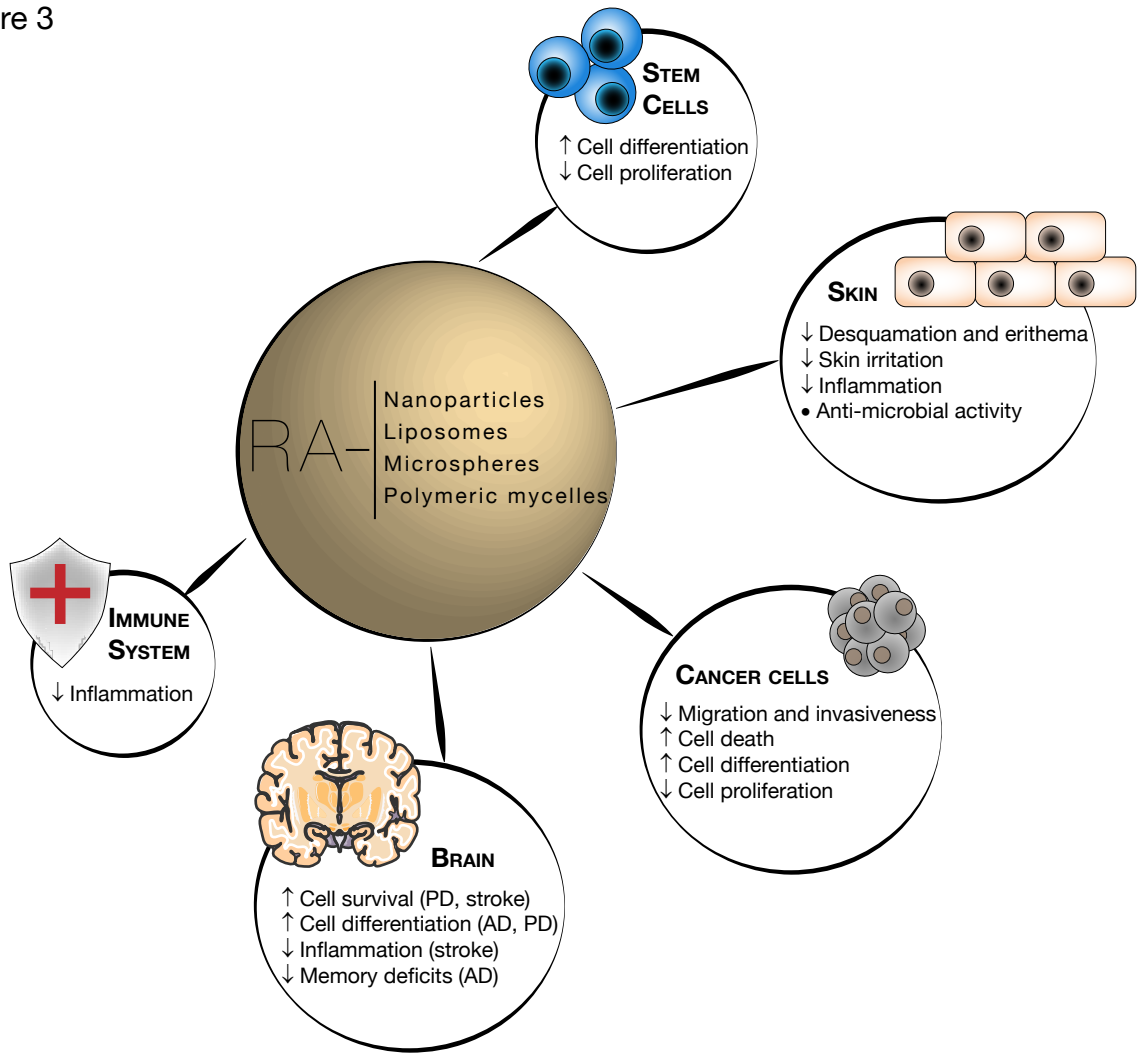
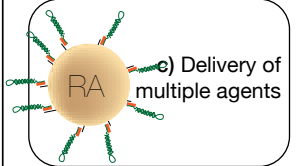
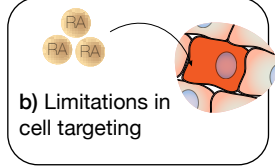
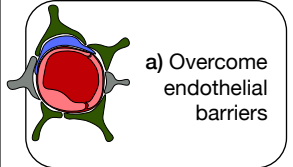


Figure 3



Challenges



Advances

